



Use of calcium in bareroot pine nurseries

David B South

College of Forestry, Wildlife and Environment, Auburn University, AL

✉ southdb@auburn.edu

Abstract

Bareroot nursery managers may apply dolomite, gypsum, or Ca-nitrate to increase Ca in nursery soils. Although a few managers follow S.A. Wilde's recommendations and maintain soil at levels of 500 to 1,000 $\mu\text{g g}^{-1}$ Ca, there is no need to keep Ca levels this high. In contrast, managers at sandy nurseries apply Ca when soil tests drop below 200 $\mu\text{g g}^{-1}$ Ca. In fact, acceptable pine seedlings have been produced in irrigated soil with <100 $\mu\text{g g}^{-1}$ available Ca. In plantations, asymptomatic wildlings grow when topsoil contains 17 $\mu\text{g g}^{-1}$ Ca. In sandy soils, applying too much gypsum can result in a temporary Mg deficiency and too much lime will result in chlorotic needles.

Managers apply Ca when foliar levels fall below a published "critical value." The belief that the critical value for Ca varies by stock type is not valid. In fact, numerous "critical" values are invalid since they were not determined using growth response curves. Critical values determined for small seedlings using CaCl_2 in sand are apparently not valid for use in bareroot nurseries.

At bareroot nurseries, the soil extractable Ca level can decline during a year by 30 $\mu\text{g g}^{-1}$ or more. Harvesting 1.7 million pine seedlings may remove 20 kg ha^{-1} of Ca but irrigation can replace this amount or more. When water contains 5 mg l^{-1} Ca, 600 mm of irrigation will add 30 kg ha^{-1} Ca. In some areas, 1,000 mm of rainfall will supply 7 kg ha^{-1} Ca. Even when a Mehlich 1 test shows no exchangeable Ca in the topsoil, pine needles on tall trees may exceed 2,000 $\mu\text{g g}^{-1}$ Ca due to root growth in subsoil.

There are few documented cases of deficient pine needles (<300 $\mu\text{g g}^{-1}$ Ca) in irrigated nurseries in Australia, New Zealand, Scotland and in the Americas. Even when soil fumigation delays the inoculation of ectomycorrhiza, bareroot pines have adequate levels of Ca. Typically, foliage samples from pine nurseries contain at least 1,000 $\mu\text{g g}^{-1}$ Ca. Samples from 9-month-old seedlings range from 300 to 11,000 $\mu\text{g g}^{-1}$ Ca. Although the "critical value" for *Pinus echinata* foliage is not known, seedlings with 300 $\mu\text{g g}^{-1}$ Ca were not stunted and apparently grew well after outplanting.

Keywords

Nutrition; Foliar analysis; Soil testing; Hidden hunger; Toxicity

Contents

1	Introduction	108
2	History	109
3	Soil tests	111
4	Tissue analysis	113
4.1	Hidden hunger	116
4.2	Critical level in foliage	117
4.3	Visible symptoms	119

ARTICLE INFO

Citation:

South DB (2022) Use of calcium in bareroot pine nurseries. *Reforesta* 14: 107-152.

DOI: <https://dx.doi.org/10.21750/REFOR.14.07.102>

Editor: Jovana Devetaković

Received: 2022-11-04

Accepted: 2022-12-19

Published: 2022-12-29



Copyright: © 2022 South David B.

This work is licensed under

a [Creative Commons Attribution 4.0 International Public License](https://creativecommons.org/licenses/by/4.0/).



5	Soils	121
5.1	Soil pH	122
5.2	Organic matter	122
5.3	Nitrogen	122
5.4	Phosphorus	124
5.5	Magnesium	125
5.6	Ca/Mg ratio	126
5.7	Manganese	126
6	Irrigation water	127
7	Mycorrhiza	128
8	Calcium removed at harvest	129
9	Calcium concentration after transplanting	129
10	Toxicity	130
11	Ca Fertilizers	130
11.1	CaCO ₃	130
11.2	Ca(OH) ₂	132
11.3	CaCO ₃ · MgCO ₃	132
11.4	CaSO ₄ · 2H ₂ O	132
11.5	Ca(NO ₃) ₂	133
11.6	CAN	134
11.7	CaCl ₂	134
11.8	Ca(H ₂ PO ₄) ₂ · H ₂ O	135
11.9	Ca(H ₂ PO ₄) ₂ · CaSO ₄	135
11.10	Chelates	136
11.11	Unknown anions	136
12	Cost	136
13	Conclusions	137
14	Acknowledgments	137
15	References	137

1 Introduction

The term calcium originates from the Latin word *calx* which means lime. Due to the quantity of limestone, calcium (Ca) is the fifth most common element in the Earth's crust and the third most common element in pine seedlings. Limestone contains calcite and aragonite which are forms of CaCO₃. When limestone contains MgCO₃·CaCO₃ it is referred to as dolomite while sources with low levels of Mg are called agricultural lime (<https://www.lime.org/about-us/faqs/>). Both types have been used in bareroot nurseries but most managers prefer to apply dolomite since it contains Mg. Details about the role of Ca in plants have been reviewed previously (Hawkesford et al. 2012; Bryson and Mills 2014; White 2015; Prado 2021). Ca moves easily with water to the top of trees and is sequestered in needles. Typically, Ca concentration in new needles increases during growing season. Since retranslocation from needles to the phloem is limited (Helmisaari 1992), concentrations of Ca in 18-month-old needles are typically higher than in 9-month-old needles. A literature review of Ca was undertaken to establish what has been learned from over a century of research and use in bareroot pine nurseries.

[Abbreviations: AA = ammonium acetate soil extraction. B = boron. Ca = calcium. CEC = cation exchange capacity. Cl = chloride. Fe = iron. K = potassium. LSD₀₅ = Least significant difference, α = 0.05. meq = milliequivalent. Mg = magnesium. Mn =

manganese. N = nitrogen. P = phosphorus. pH = potential hydrogen. Pt = *Pisolithus tinctorius*. S = sulfur. TSP = triple superphosphate. Soil pH was measured in water.]

2 History

Most nursery managers in the 19th century did not apply fertilizers but a few spread lime before sowing cover-crops. At new nurseries located on nutrient-poor soils, organic matter was incorporated into soil before sowing seed and sometimes lime was added to assist in decomposition (Nicol 1820). However, on new land with no organic amendments, adding lime was found to do more harm than good (Nicol 1820).

Before 1892, agricultural experiment stations analyzed commercial fertilizers for N, P and K content but not for Ca content. At that time the terms “lime” and “calcium” were synonyms. In one bulletin, the words “lime”, “gypsum” and “calcium” were mentioned 11, 2 and zero times, respectively (Scovell 1890). Fertilizer sold as raw bone manure might have an estimated nutrient value (1890) of \$0.038 kg⁻¹.

Schenck (1907) realized Ca was an essential element and he said pine seedlings contained about 19 kg Ca ha⁻¹. To replenish nutrients at his nursery, Schenck applied several fertilizers including bone meal (12% Ca). However, he admitted that results from tests with bone meal were uncertain. Although Schenck did not mention rate, 112 kg ha⁻¹ of bone meal was applied at the Greenwood Nursery (PA) (Retan 1914). At two nurseries, he found that acid phosphate (336 kg ha⁻¹) was the best fertilizer tested. In addition, sodium nitrate (168 kg ha⁻¹) and potash (168 kg ha⁻¹) were applied before sowing a cover crop.

In New York, managers believed that seedbeds of *Pinus strobus* “must be heavily fertilized but they should not be located on areas that have been heavily limed” (Steer 1915). The practice then was to apply fertilizers before sowing a cover-crop. Managers at the Syracuse Nursery (NY) applied sodium nitrate, potash, and bone meal, each at 140 kg ha⁻¹. In addition, a dilute phosphoric acid solution that was buffered with sodium, magnesium, and calcium salts (acid phosphate) was applied at 280 kg ha⁻¹.

Toumey (1916) also discussed nursery fertilization but did not mention calcium. For example, the term “phosphates of lime” (Scovell 1890, Fox 1904) means calcium phosphate. Toumey’s list of chemical fertilizers included gypsum, lime, clay and sand. He said these four substances were “chiefly valuable in improving the physical qualities of certain soils.”

Tillotson (1917) discussed the use of lime and fertilizers in Federal nurseries. He used the word “lime” 30 times and did not mention “calcium.” He said that lime helps to decompose organic matter and “makes the soil lose and friable.” Soil testing was considered expensive and inconvenient and test results were only good for only a few months. He said unslaked lime contains 90 to 95% lime and is generally used as part of a compost. In Minnesota, Hansen (1923) saw no consistent advantage of applying 4,000 kg ha⁻¹ of lime one week before sowing pine seeds.

Jones (1925) said the life of the Savanac Nursery depended upon soil fertility and the fertilizers used were mainly concentrated commercial fertilizers. Blood meal (2,000 kg ha⁻¹) was used as a N source and bone meal (1,000 kg ha⁻¹) was used as a P source. Hydrated lime (3,900 kg ha⁻¹) was applied at the Savanac Nursery in order to increase soil pH (Jones 1925). Hydrated lime is calcium hydroxide [Ca(OH)₂] while dehydrated lime [CaO] is known as quicklime (Table 1).

Wahlenberg (1930) installed various fertilizer trials in hopes of advancing nursery management. In one trial he tested six sources of calcium. The best germination

of pine occurred with calcium monohydrogen phosphate (355 kg ha⁻¹). He might have not been the first to test gypsum on pine seedbeds but he may have been the first in North America to publish test results. In California, several experiments were conducted to improve nursery management (Show 1930). At one nursery, fertilization was deemed unnecessary but at another, liming increased root mass of pine by more than 100%.

Wakeley (1935) said that fertilization of southern pine nursery had just begun and practically no fertilizer trials had been established. He mentioned N, P, K, bone meal and lime, but did not mention calcium. Managers realized that too much lime could increase damping-off fungi (Wilde 1938; Auten 1945; Stoeckeler 1949) and, therefore, applying lime was generally avoided in pine nurseries (Wilde 1942). When lime was recommended for pine seedbeds, the target acidity was generally pH 4.8 (Wilde 1942). Several university laboratories tested soil acidity before 1940 (Lunt 1938; Wilde 1938; Flaten 1939; Argetsinger 1941).

Table 1. A partial list of calcium (Ca) fertilizers. Powder (P): granules (G): liquids (L).

Ingredients	Name	Form	% Ca	% Other
Calcium oxide	Quicklime	P	65	
Calcium hydroxide	Slacked lime	P	54	
Calcium carbonate	Ag lime	G	32	3 Mg
Calcium chloride	De-ice	G	36	64 Cl
Phosphorite	Rock phosphate	G	33	33 P
Calcium sulfate	Gypsum	G	23	18 S
Calcium magnesium carbonate	Dolomite	G	22	11 Mg
Calcium nitrate	15.5-0-0	G	19	15.5 N
Calcium phosphate + CaSO ₄	0-20-0	G	19	8 P 11 S
Calcium phosphate	0-45-0	G	15	19 P
Calcium magnesium nitrate	13.5-0-0	G	12	13 N 4 Mg
Calcium ammonium nitrate	17-0-0	L	9	17 N
Calcium EDTA	Calcium chelate	P	9	3 N
Calcium ammonium nitrate	27-0-0	G	8	27 N
Calcium glucoheptonate	Tracite GHEP	L	8	
Urea and CaCl ₂	23-0-0	L	7	14 Cl
Limestone ammonium nitrate	LAN	G	4	28 N 2 Mg

In the 20th century, there were two schools of thought regarding how much Ca should be added to nursery soils. The “experiment” school established fertilizer trials while the “short-cut” school guessed at fertility needs of conifers and hardwoods. Making nursery recommendations base on fertility tests of virgin forest soils took less time than installing and measuring nursery fertility trials (Wilde 1940). As a result, Wilde set high Ca target levels for nursery soil and varied fertility targets by species. His soil Ca targets were 500, 750, and 1,250 µg g⁻¹ for *Pinus banksiana*, *Pinus resinosa* and *Pinus strobus*, respectively (Wilde 1938). Once he published tentative standards for conifers in nurseries, many accepted these “short cut” standards without asking about the benefit/cost ratio. As a result, individuals likely applied Ca to *Pinus strobus* seedbeds while applying no Ca in adjacent seedbeds sown with *Pinus banksiana*. The belief that pine seedbeds should contain 500 to 1,000 µg g⁻¹ Ca continued into the 21st century (Briggs 2008; VandeHey 2007; Bueno et al. 2012). In contrast, pines can grow well in irrigated nursery soil with >50 µg g⁻¹ available Ca (Table 2) and in non-irrigated sites when soil contains 30 µg g⁻¹ available Ca (NCSFNC 1991).

Table 2. Height (Ht), root-collar diameter (RCD), green root mass (Root), green total mass (Total) and % short roots with ectomycorrhiza (Ecto) for seedlings growing in sandy soil with less than 140 $\mu\text{g g}^{-1}$ calcium (Mehlich 1). Soil samples were collected before sowing (Marx et al. 1984). Nurseries are arranged in order of increasing soil Ca at time of sowing.

Species	Nursery	State	pH	Sand	Ca	Ht	RCD	Root	Total	Ecto
				%	$\mu\text{g g}^{-1}$	cm	mm	g	g	%
<i>Pinus taeda</i>	Champion	SC	5.1	88	56	21.4	4.9	3.7	12.9	20
<i>Pinus taeda</i>	Westvaco	SC	4.7	86	80	26.6	7.9	13.0	34.4	61
<i>Pinus taeda</i>	Weyerhaeuser	OK	5.2	89	89	20.3	3.9	2.4	11.4	37
<i>Pinus elliotii</i>	Buckeye	FL	4.7	88	91	22.6	4.8	1.8	13.5	23
<i>Pinus resinosa</i>	Toumey	MI	4.7	86	96	21.4	4.7	5.4	26.3	44
<i>Pinus elliotii</i>	Buckeye	FL	5.0	94	101	27.0	4.5	2.3	13.9	27
<i>Pinus taeda</i>	Weyerhaeuser	OK	4.7	91	103	24.1	4.8	3.0	14.7	18
<i>Pinus virginiana</i>	Vallonia	IN	4.3	82	105	20.8	4.5	8.9	21.8	19
<i>Pinus taeda</i>	Weyerhaeuser	OK	5.5	90	106	28.2	4.4	3.4	14.0	43
<i>Pinus resinosa</i>	USDA	MI	4.8	88	114	14.1	4.1	2.5	13.1	46
<i>Pinus virginiana</i>	Vallonia	IN	5.1	81	131	25.1	4.0	5.1	14.9	43
<i>Pinus elliotii</i>	Buckeye	FL	5.2	95	139	19.1	3.0	1.2	6.6	28

The experiment school of thought was led by researchers in Germany, New Zealand and the United Kingdom who based fertility recommendations on nursery experiments. German researchers believed guessing at Ca needs for pine was not as useful as fertilizer tests. In one experiment in 1925, liming soil (5,400 Mg ha⁻¹) resulted in a statistically significant reduction in seedling numbers (Steven 1928). Likewise, too much Ca acetate reduced growth of *Pinus echinata* (Chapman 1941).

3 Soil tests

Soils contain exchangeable and non-exchangeable Ca and the sum is referred to as total soil calcium. In some soils, the total may be 14,000 $\mu\text{g g}^{-1}$ Ca while the exchangeable portion is 6,090 $\mu\text{g g}^{-1}$ Ca (Hallett and Hornbeck 1997). Various methods (Mehlich 1, Mehlich 3, AA, etc.) are used to estimate the exchangeable amount (Alva 1993; Davey 2002; Mylavarapu et al. 2002). Extractions of identical soil samples might produce 100 $\mu\text{g g}^{-1}$ Ca when using Mehlich 1 and 170 $\mu\text{g g}^{-1}$ Ca when using Mehlich 3 (Mylavarapu et al. 2002). As a result, managers who use Mehlich 1 will likely apply more Ca fertilizers than those who rely on Mehlich 3 tests.

In theory, the Mehlich 3 tests should not vary among laboratories, but that is not the case. In one comparison, laboratory C extracted twice as much Ca as did laboratory B (Table 3). Therefore, those who use laboratory C will apply less Ca to their seedbeds than managers who send samples to laboratory A or B.

Table 3. Examples of calcium soil test results (Mehlich 3) using three soil samples. Two laboratories produced similar results but laboratory C extracted twice as much calcium than laboratory B.

Sample	Laboratory		
	A	B	C
	$\mu\text{g g}^{-1}$	$\mu\text{g g}^{-1}$	$\mu\text{g g}^{-1}$
9	63	93	308
12	71	89	191
16	49	102	203

When a soil test value is below a pre-determined “trigger,” then managers usually incorporate Ca several months before sowing pine seed. Due to equipment and budget limitations, a “stair-step” recommendation curve may be adopted (Figure 1). When a soil test indicates low Ca at pH 5, most agronomists recommend lime but nursery managers usually apply gypsum since pines grow better in acid soils (Marx 1990; South 2017). Gypsum may be applied at 363 kg ha⁻¹ (Sonne 2006), or 900 kg ha⁻¹ or 1,500 kg ha⁻¹. In one nursery trial, gypsum (7,500 kg ha⁻¹) applied before sowing did not increase growth of *Pinus radiata* seedlings (Flinn and Waugh 1983).

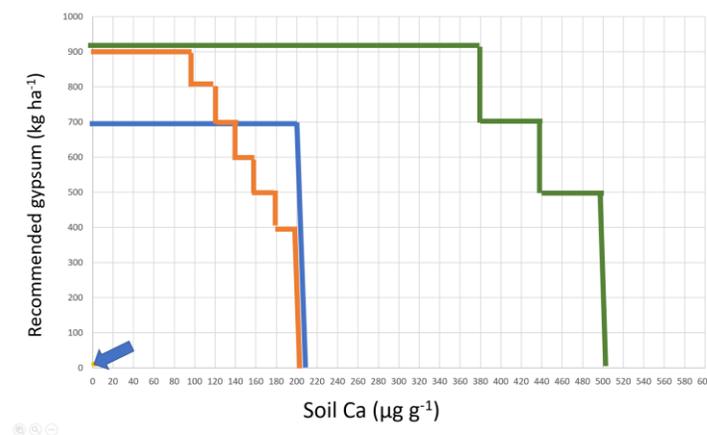


Figure 1. Various opinions exist regarding calcium (Ca) fertilization of pine seedbeds (White et al. 1980). Some researchers set 200 $\mu\text{g g}^{-1}$ Ca as a satisfactory level (Woodwell 1958; South and Davey 1983) while others use a 500 $\mu\text{g g}^{-1}$ (Briggs 2008; Bueno et al. 2012) or 600 $\mu\text{g g}^{-1}$ target (van den Driessche 1984). When a Mehlich 3 test indicates a soil (pH 5.5) contains 165 $\mu\text{g g}^{-1}$ Ca, one manger might apply 500 kg ha⁻¹ of gypsum (orange line), another might apply 700 kg ha⁻¹ (blue line) and a third might apply 900 kg ha⁻¹ (green line). In contrast, some agronomists recommend no gypsum (arrow) and instead recommend raising soil pH by applying various rates of dolomitic lime.

Nursery managers apply Ca either from experience or from the recommendations of others. Some managers do not apply Ca since irrigation supplies more than required (Armson and Sadreika 1979; Argo et al. 1997). At nurseries where irrigation water is below 20 $\mu\text{g g}^{-1}$ Ca, most managers apply the same rate of Ca to all pines even though Wilde (1938) recommend applying more Ca to *Pinus banksiana* than to *Pinus resinosa*. His logic for fertilizing with high rates of Ca included soil factors that did not relate to seedling nutrition (Wilde 1946, p. 196). In contrast, others recommend lower Ca levels which reduces expenditures for gypsum (Table 4).

Table 4. Recommended levels of extractable calcium for growing pine seedlings in bareroot nurseries vary by individual. Wilde (1938) and Stoeckeler and Jones (1957) varied the level by species and May (1984) varied it by sand content. Some values in this table were “tentative” and were subject to modification (Solan et al. 1979).

Species or soil texture	Minimum $\mu\text{g g}^{-1}$	Recommended $\mu\text{g g}^{-1}$	Upper value $\mu\text{g g}^{-1}$	Reference
--	100	-	300	Sadreika 1976
--	200	-	-	South and Davey 1983
Sand	200	-	300	May 1984
--	-	300	-	Hallet 1980
--	-	300	-	Bunting 1980
--	350	-	400	Kormanik et al. 1994
Sandy loam	300	-	450	May 1984
--	125	375	500	Knight 1978a
<i>Pinus banksiana</i>	-	400	-	Stoeckeler and Jones 1957
<i>Pinus banksiana</i>	-	500	-	Wilde 1938
--	-	500	-	Solan et al. 1979
<i>Pinus sylvestris</i>	-	600	-	Stoeckeler and Jones 1957
--	-	600	-	van den Driessche 1984
--	500	-	1,000	Youngberg 1984
<i>Pinus resinosa</i>	-	750	-	Wilde 1938
--	-	1,000	-	Landis 1988
<i>Pinus strobus</i>	-	1,250	-	Wilde 1938

4 Tissue analysis

Soil analyses are used to determine how much lime and gypsum to apply in bareroot nurseries (before sowing) while tissue analyses are used to diagnose deficiency and monitor uptake of nutrients. Except when extractable Ca is below $5 \mu\text{g g}^{-1}$ Ca (e.g. Chaves and Corrêa 2005), there typically is no relationship between soil Ca and foliar Ca concentration (South et al. 2017; Figure 2).

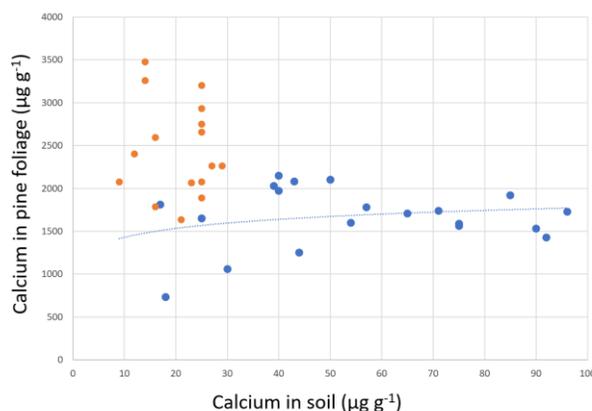


Figure 2. Irrigated *Pinus elliottii* seedlings in a greenhouse (16 orange dots; Steinbeck 1962) and non-irrigated *Pinus taeda* trees in plantations (20 blue dots and blue line; NCSFNC 1991) show no relationship between soil calcium and foliar calcium (blue $r = 0.22$). *Pinus elliottii* seedlings were irrigated using water containing $7.9 \mu\text{g g}^{-1}$ calcium while no irrigation was applied to the *Pinus taeda* plantations.

In nurseries, Ca in irrigation water can explain the lack of a relationship while root growth deeper than 15 cm may explain no relationship in pine plantations. Even when extractable soil Ca in the top 60 cm is below detectable levels (Mehlich 1), *Pinus palustris* trees with deep root may contain 2,900 Ca $\mu\text{g g}^{-1}$ in needles (McLeod et al. 1979). In contrast, with lime rate studies, there can be a high correlation between foliar Ca concentration and soil Ca (Carter 1987).

Pinus taeda seedlings with less than 1,000 $\mu\text{g g}^{-1}$ Ca can be found in the southern United States (Table 5) while seedlings with more than 6,000 $\mu\text{g g}^{-1}$ Ca in foliage occur in the Inland Mountain West (Mexal and Fisher 1987; Landis 1988). Although some claim the range of adequate % foliar Ca is greater for bareroot pines (versus container stock), data do not support the belief that bareroot pine foliage should contain 1,000 $\mu\text{g g}^{-1}$ more Ca than foliage sampled from container-grown pines (Figure 3).

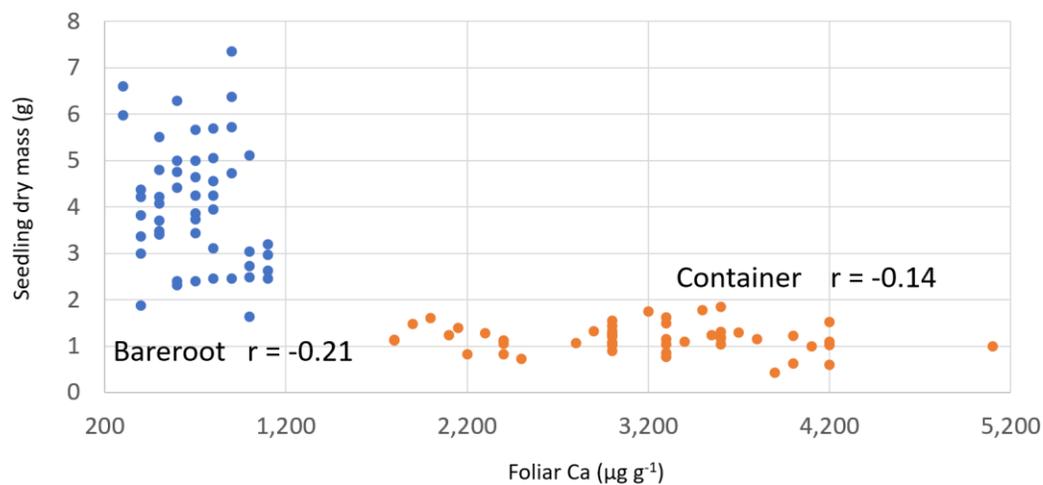


Figure 3. Seedling dry mass and calcium (Ca) concentration in foliage for 50 *Pinus echinata* seedlings (orange dots) grown in a greenhouse at the Stephen F. Austin State University (Bryson 1980). Blue dots represent 50 bareroot *Pinus echinata* seedlings lifted in November 1978 at the Ashe Nursery. There was no relationship ($\alpha = 0.10$) between foliar calcium (Ca) and seedling dry mass for either stock type.

In theory, Ca concentrations in pine needles should increase with seedling age. For example, *Pinus taeda* seedlings in August contained 3,050 $\mu\text{g g}^{-1}$ Ca while six months later the average was 3,750 $\mu\text{g g}^{-1}$ Ca (Danielson 1966). Likewise, *Pinus radiata* needles sampled in fall may be 1,000 $\mu\text{g g}^{-1}$ Ca lower than needles collected three months later (Menzies et al. 1981; Olykan et al. 1995). However, there are exceptions to theory. At some nurseries there is no trend (Sung et al. 1997) and sometimes June values were higher than concentrations in September (May et al. 1962; Baer 1984).

Table 5. The foliar calcium concentration of pine seedlings. Data compiled from Voigt 1955; Voigt 1958; Fowells and Krauss 1959; May et al. 1962; Steinbeck 1962; Beaton et al. 1965; Steinbeck 1965; Danielson 1966; Metz et al. 1966; Malavolta et al. 1970; Iyer et al. 1971; van Lear and Smith 1972; Woessner et al. 1975; Knight 1978b; Landis 1979; Hart and Widdowson 1981; Bryson 1980; Flinn et al. 1980; Baer 1984; van den Driessche 1984; Boyer and South 1985; Richardson and Perkins 1985; Landis 1988; Gleason 1989; Wall 1994; Andrews et al. 1998; Jose et al. 2003; Leski et al. 2010; Starkey and Enebak 2012; Hans 2013; Januszek et al. 2014.

Species	Year-Month	samples	Foliar calcium ($\mu\text{g g}^{-1}$)			Reference
			Mean	Minimum	Maximum	
<i>Pinus banksiana</i>	1948?	5	2,740	2,000	3,300	1958
	1969?	5	2,300	2,000	2,700	1971
<i>Pinus caribaea</i>	1979-6	21	1,945	1,300	3,600	1981
<i>Pinus contorta</i>	1961-1963	34	3,960	1,600	6,400	1965
	1968-78	53	3,200	1,900	5,200	1984
<i>Pinus echinata</i>	1978-11	50	724	300	1,100	1980
<i>Pinus elliotii</i>	1954-12	39	1,490	900	2,220	1962
	1959-6	54	3,510	1,860	7,920	1962
	1959-9	54	850	500	1,800	1962
	1959-12	54	2,460	1,720	3,580	1962
	1970	4	4,550	3,800	5,000	1970
	1969	12	9,460	4,100	19,200	1972
	1961-12	48	2,224	1,450	3,450	1962
<i>Pinus palustris</i>	2000-10	8	4,675	3,000	7,400	2003
<i>Pinus ponderosa</i>	1978-11	2	6,700	--	11,300	1979
	1979-5	9	3,060	1,200	5,000	1984
	1980-4	9	2,760	1,100	3,500	1984
	1981-4	9	5,230	1,300	9,500	1984
	1985-9	9	2,840	2,270	3,510	1989
<i>Pinus radiata</i>	1953?	17	5,465	1,900	7,600	1955
	1975	18	5,100	2,200	5,200	1978
	1971-1975	12	3,300	1,700	5,300	1980
	1983-4	7	3,330	1,700	5,800	1985
	2011-8	22	1,960	1,300	2,600	2013
<i>Pinus resinosa</i>	1969?	7	2,500	1,900	3,500	1971
<i>Pinus strobus</i>	1998?	78	5,000	3,000	8,000	1998
<i>Pinus sylvestris</i>	1963-1	45	4,130	2,200	7,700	1965
	2003-11	15	4,660	3,300	5,800	2014
	2006-9	15	4,260	3,600	4,800	2010
<i>Pinus taeda</i>	1955-1	162	2,130	1,430	3,830	1962
	1956-1	216	1,370	810	2,480	1962
	1957-12	12	1,420	900	2,500	1959
	1962-2	256	1,200	340	2,160	1966
	1965-8	4	3,050	2,700	3,400	1966
	1966-2	4	3,650	3,600	3,700	1966
	1973-4	19	1,384	800	2,000	1975
	1982-12	41	3,000	2,200	6,600	1985
	1989-1	5	3,780	3,600	4,100	1994
	2009-7	19	3,300	2,900	5,500	2012
	2009-10	19	3,500	2,100	4,800	2012
<i>Pinus virginiana</i>	2010-2	19	3,300	2,500	5,900	2012
	1957-12	12	2,030	1,600	2,500	1959

4.1 Hidden hunger

Since a hidden hunger for Ca might not occur in 99.9% of pine plantations (Snowdon and Waring 1985; Will 1985; Woollons et al. 1995; May et al. 2009), there is little need to fertilize pine plantations with Ca. Most researchers in Australia and North America do not recommend adding lime or gypsum to pine stands at planting or at mid-rotation. In Europe, lime may be applied in forests (Baule 1975), but this is due mainly to increase soil pH as part of a precautionary principle (Talkner et al. 2019). Growth gains from applying dolomite to a 9-year-old *Pinus taeda* plantation (pH 4.3) were likely due to a Mg-deficiency ($<700 \mu\text{g g}^{-1}$ Mg in foliage) since foliage in control plots contained $>1,100 \mu\text{g g}^{-1}$ Ca (Kyle et al. 2005). No hidden hunger was detected when a nursery soil contained $135 \mu\text{g g}^{-1}$ Ca (South et al. 2017) or when a silt loam soil contained $246 \mu\text{g g}^{-1}$ Ca (McKee 1978). In fact, no hidden hunger was detected when a piedmont soil contained $57 \mu\text{g g}^{-1}$ Ca (Moschler et al. 1970). In contrast, *Fraxinus pennsylvanica* exhibited a hidden hunger for Ca at a bareroot nursery in North Carolina (Deines 1973).

Some surveys suggest a positive relationship between foliar Ca and height growth of pines (Hoyle and Mader 1964; Jenkinson 1974) while others report a negative relationship (Wells et al. 1973). A cause-and-effect relationship is unlikely since these correlations are confounded with other nutrients. Likewise, a positive correlation between soil Ca and height growth of *Pinus taeda* in a bareroot nursery (South et al. 2018) does not prove seedlings have a hidden hunger for Ca.

A hidden hunger can be proved by establishing a rate trial and plotting a response curve. Response curves using nutrient-sand culture were developed by varying CaCl_2 (Mitchell 1938; Woodwell 1958). In one study (Figure 4), a 10% reduction in maximum growth occurred at about $80 \mu\text{g g}^{-1}$ soil Ca and $2,700 \mu\text{g g}^{-1}$ foliar Ca (Mitchell 1939). For *Pinus serotina* and *Pinus taeda*, response curves were nearly flat (Woodwell 1958). However, a response curve developed in a greenhouse using washed sand and deionized irrigation water may not be appropriate for use in an operational nursery (Bates 1971). In a greenhouse, good root growth of *Pinus taeda* occurred at $22 \mu\text{g g}^{-1}$ soil Ca (Lyle and Adams 1971) and good shoot growth occurred at $20 \mu\text{g g}^{-1}$ soil Ca (Woodwell 1958).

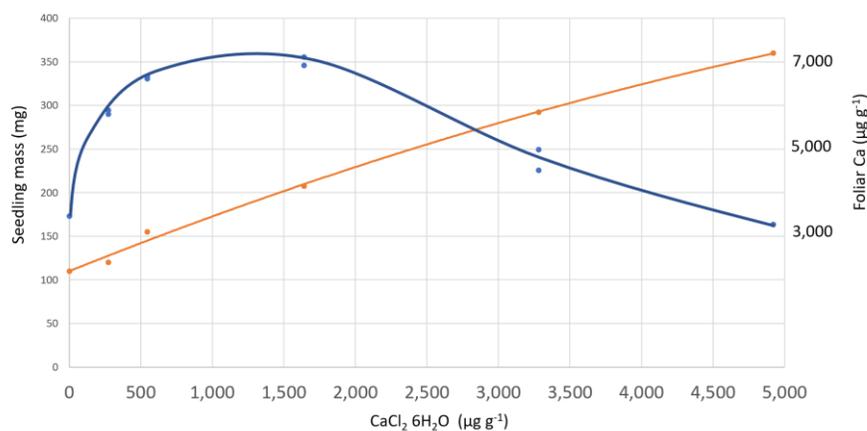


Figure 4. The effect of calcium chloride on dry mass (blue line) and foliar calcium concentration (orange line) of *Pinus strobus* seedlings growing in sand at the Black Rock Experimental Nursery (Mitchell 1939). Solution concentrations of $100 \mu\text{g g}^{-1}$ calcium (Ca) and $200 \mu\text{g g}^{-1}$ chloride (Cl) are equivalent to $546 \mu\text{g g}^{-1}$ $\text{CaCl}_2 \cdot 6\text{H}_2\text{O}$. Seedlings with a 315 mg mass likely had $2,700 \mu\text{g g}^{-1}$ Ca in needles.

Due to fertility management and irrigation in nurseries, bareroot pine seedlings likely have flat response curves for Ca. As a result, pine seedlings can grow well at irrigated nurseries where the extractable soil Ca is 50 to 150 $\mu\text{g g}^{-1}$ (Table 2). Even in a greenhouse, flat response curves have been observed in pots filled with sand (Steinbeck 1962; Figure 5). A growth response might result at locations where extractable soil Ca is below 5 $\mu\text{g g}^{-1}$ (e.g. Chaves and Corrêa 2005).

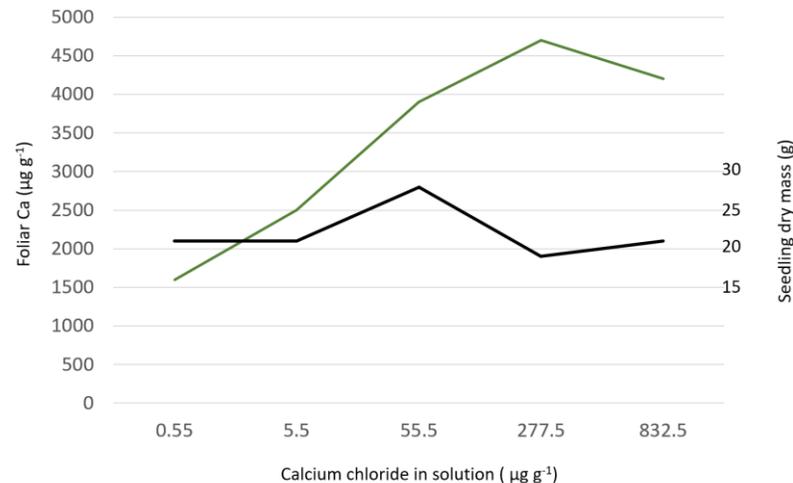


Figure 5. The effect of calcium chloride on dry mass (black line) and foliar calcium concentration (green line) of *Pinus virginiana* seedlings growing in sand in a greenhouse (Sucoff 1962). Due to unmeasured Ca in water, it was assumed that the solution concentration needed for good growth was 3 $\mu\text{g g}^{-1}$ Ca. The nutrient solutions also contained 120 $\mu\text{g g}^{-1}$ of magnesium sulfate as well as K, Na, N and P. Ca deficiency symptoms did not occur in any treatment. Increasing the concentration of CaCl_2 increase Ca in needles ($\text{LSD}_{05} = 2,500 \mu\text{g g}^{-1}$ Ca) but did not affect seedling dry mass ($\text{LSD}_{05} = 11$ g).

4.2 Critical level in foliage

The “critical level” of foliar Ca is defined as the concentration that occurs at 90% of maximum yield; the probability of a growth response to fertilization is high when foliage is below this level (Bates 1971; Kumar and Shivay 2008). Most critical values for Ca in pine are guesses since the values were not determined from response curves. I see no reason to manage pine nurseries based on assumptions made using Ca ranges derived from foliage sampled from pine stands. Therefore, an accurate critical value for Ca has not been determined for *Pinus taeda*. During the 20th century, there was insufficient information to predict a pine response to Ca fertilization (Stone 1953; Allen 1987; Will 1985) and this situation has not changed. Although critical-values for pine foliage have been repeated numerous times, these values were not independently verified. Perhaps one estimate was equated with the average foliar Ca value for *Pinus taeda* (Metz et al. 1966). A “critical level” of 500 $\mu\text{g g}^{-1}$ Ca for *Pinus ponderosa* was likely based on the optimum N/Ca ratio for *Betula verrucosa* (Ingestad 1974). Without complete explanations, we are left wondering what procedures were used to determine most critical values listed in Table 6.

When bareroot pines contained 800 $\mu\text{g g}^{-1}$ Ca (Figure 3), I doubt growth in 1978 would have increased after seedlings received a foliar application of CaCl_2 . I contend that for pine, critical levels >500 $\mu\text{g g}^{-1}$ Ca (Table 6) can be ignored. Before managers

top-dress pine seedlings based on foliar values of 2,000 $\mu\text{g g}^{-1}$ Ca, researchers should first install tests to determine if bareroot seedlings with 800 $\mu\text{g g}^{-1}$ Ca in foliage will respond to applications of CaCl_2 .

Table 6. Estimated critical values for pine needles vary by species and reference. None of these values have been verified using Ca trials in bareroot nurseries.

Species	$\mu\text{g g}^{-1}$ Ca	Method used	Reference
<i>Pinus resinosa</i>	350	Cited Swan 1972	Timmer and Armstrong 1987
<i>Pinus spp.</i>	500	Not provided	Zöttl 1973
<i>Pinus caribaea</i>	500	Cited Zöttl 1973	Vettorazzo and Couto 1997
<i>P. ponderosa + P. jeffreyi</i>	500	Cited Ingestad 1974	Powers 1983
<i>Pinus ponderosa</i>	500	Not provided	Allen 1987
<i>Pinus ponderosa</i>	500	Cited Powers 1983	Moore et al. 2004
<i>Pinus radiata</i>	600	Cited others	Boardman et al. 1997
<i>Pinus contorta</i>	800	Cited Swan 1972	Moore et al. 2004
<i>Pinus contorta</i>	800	Not provided	Binkley and Fisher 2013
<i>Pinus contorta</i>	800	Not provided	Allen 1987
<i>Pinus elliotii</i>	800	Not provided	Pritchett and Comerford 1981
<i>Pinus sylvestris</i>	900	Greenhouse data	Hacskaylo et al. 1969
<i>Pinus sylvestris</i>	900	Cited van den Burg 1990	Mellert and Göttlein 2012
<i>Pinus banksiana</i>	1,000	Response curve	Swan 1970
<i>Pinus elliotii</i>	1,000	Not provided	Allen 1987
<i>Pinus radiata</i>	1,000	Not provided	Knight 1978
<i>Pinus palustris</i>	1,000	Cited Blevins et al. 1996	Dickens et al. 2021
<i>Pinus elliotii</i>	1,200	Cited others	Boardman et al. 1997
<i>Pinus taeda</i>	1,200	Not provided	Allen 1987
<i>Pinus taeda</i>	1,200	Not provided	Gregoire et al. 2004
<i>Pinus elliotii</i>	1,400	Greenhouse data	Malavolta et al. 1970
<i>Pinus taeda</i>	1,500	Not provided	Jokela 2004
<i>Pinus sylvestris</i>	2,000	Cited Göttlein 2015	Talkner et al. 2019
<i>Pinus strobus</i>	2,700	Response curve	Mitchell 1939
<i>Pinus nigra</i>	2,700	Response curve	Kavvadias and Miller 1999
<i>Pinus sylvestris</i>	3,400	Response curve	Kavvadias and Miller 1999

When foliar Ca values fall below an accurate critical value and plants are in the “hidden hunger” zone (Mitchell 1939; Ritchey et al. 1982; Gregoire and Fisher 2004), then Ca fertilization will increase height growth. For example, when growing in acid-washed sand that was fertilized with N, seedlings with less than 80 $\mu\text{g g}^{-1}$ Ca in foliage grew taller when fertilized with CaCl_2 (Figure 6).

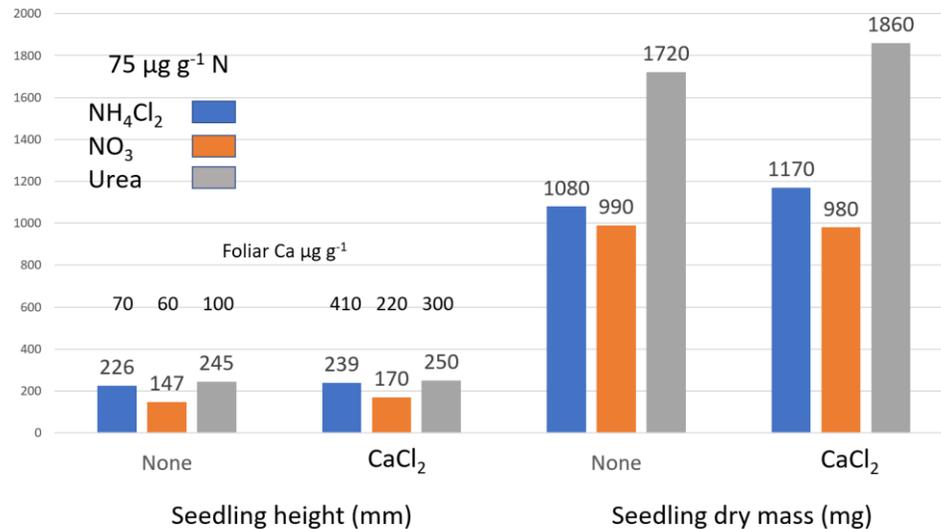


Figure 6. In a factorial trial, 18 pots containing *Pinus taeda* seedlings were fertilized with calcium chloride ($200 \mu\text{g g}^{-1} \text{Ca}$) and the remaining 18 pots (three pots per bar) were not fertilized with calcium (Pharis et al. 1964). Seedlings were fertilized three times a week using solutions of NH_4Cl_2 or urea or NO_3 (NaNO_3 then KNO_3). The CaCl_2 treatment increased height ($\alpha = 0.05$) and foliar Ca ($\alpha = 0.01$) but did not increase seedling mass ($\alpha = 0.05$). Photos of seedlings represented by this graph appear on page 563 (Pharis et al. 1964).

4.3 Visible symptoms

When Ca-deficient seedlings are produced in greenhouses, symptoms vary with species. For *Pinus taeda* (Figure 7), a Ca deficiency caused resin to exude on needles and terminal buds (Lyle 1969). *Pinus taeda* needles were yellow with yellow-red splotches along the needle while *Pinus palustris* needles were delicate and had a pale green color (Pessin 1937). *Pinus sylvestris* seedlings had dead terminal buds and beads of resin (Figure 8). Beads of resin also formed on the tips of bracts surrounding the terminal bud of *Pinus radiata* (Purnell 1958). Ca-deficient *Pinus banksiana* had beads of exuded resin and contorted needles (Swan 1970). In regards to *Pinus radiata* plantations in South Wales, Australia, Humphreys (1964) said “One of the most important symptoms which we use is the appearance of a small resin drop on the young developing needles. This can be found by examining the plants with a hand lens.” The lowest foliar Ca values reported for pine needles are $60\text{--}110 \mu\text{g g}^{-1}$ (Goslin 1959; Pharis et al. 1964; Chaves and Corrêa 2005) and these pines had visible deficiency symptoms.

Some say visual symptoms in Ca-deficient pine plantations include dieback of stem tips and branches and needle tortuosity (Rocha et al. 2019). A lack of photos illustrating these symptoms suggests documented cases of Ca-deficiencies in pine plantations are very rare (Bengtson 1968). Several traditional assumptions about Ca-deficiency are not valid (Saure 2014).

There are numerous photos of Ca-deficient crops but few if any photos illustrating symptoms on pine in irrigated bareroot seedbeds. In addition, there are no photos of Ca-deficient pines in either New Zealand (Will 1985; Davis et al. 2015), United Kingdom (Binns et al. 1980) or the United States. In contrast, photos from greenhouse trials have been published (Table 7).



Figure 7. *Pinus taeda* seedlings grown water culture in a greenhouse (photo taken August 1954 by Jack May). The Ca-deficient seedling had a dead terminal with some brown needles.

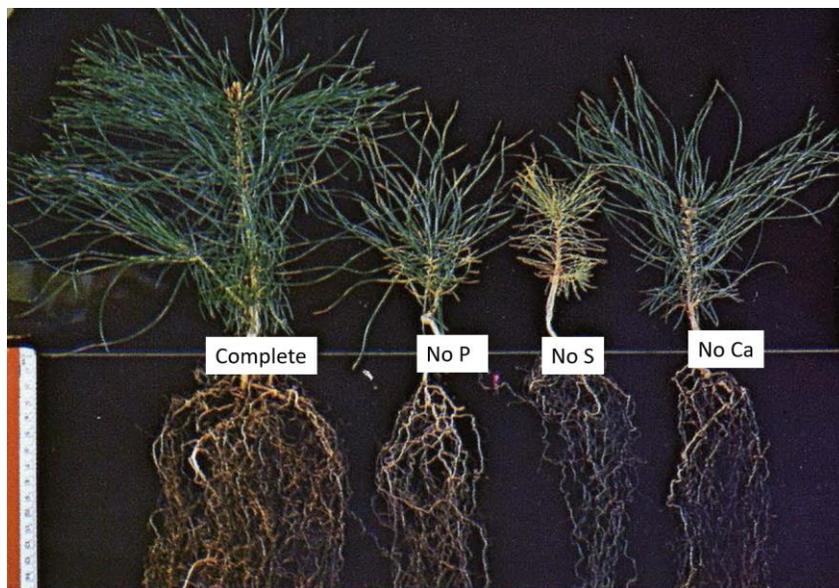


Figure 8. Seedlings of *Pinus sylvestris* were grown in sand in a greenhouse (HacsKaylo et al. 1969). Foliage sampled from complete nutrients, No P, No S and No Ca treatments were 4,800, 4,800, 6,300 and 900 $\mu\text{g g}^{-1}$ Ca, respectively. Seedlings showing Ca deficiency symptoms had dead terminals. Growth of seedlings fertilized with no Ca was greater than seedlings not fertilized with N, P or S. Roots with no added Ca were exposed to MgCl_2 , MnCl_2 , ZnCl_2 and CuCl_2 (Photo permission by The Ohio State University Extension Publishing).

Table 7. A list of references that include photographs of calcium deficiencies in greenhouse-grown pines. Foliar concentrations of Ca-deficient seedlings were included in a few references.

Species	Medium	Photo on page	More than two rates?	Reference
<i>Pinus banksiana</i>	Sand	33	Yes	Swan 1970
<i>Pinus banksiana</i>	Perlite	152	No	Donald 1991
<i>Pinus elliottii</i>	Sand	27	No	Truman 1972
<i>Pinus monticola</i>	Sand	96	No	Murison 1960
<i>Pinus nigra</i>	Perlite	508	Yes	Kavvadias 1996
<i>Pinus ponderosa</i>	Sand	96	No	Murison 1960
<i>Pinus radiata</i>	Sand	86	No	Purnell 1958
<i>Pinus sylvestris</i>	Sand	47	No	Goslin 1959
<i>Pinus sylvestris</i>	Sand	51	No	Hacskaylo et al. 1969
<i>Pinus sylvestris</i>	Perlite	508	Yes	Kavvadias 1996
<i>Pinus taeda</i>	Sand	13	No	Sucoff 1961
<i>Pinus taeda</i>	Sand	562	No	Pharis et al. 1964
<i>Pinus taeda</i>	Water	Plate 18	No	Bengtson 1968
<i>Pinus taeda</i>	Sand	32	No	Truman 1972
<i>Pinus virginiana</i>	Sand	3	Yes	Sucoff 1962

5 Soils

Bareroot nurseries in the United States contain more than 40 $\mu\text{g g}^{-1}$ Ca and those located on clay loam and sandy loam soils have more than 100 $\mu\text{g g}^{-1}$ of exchangeable Ca in topsoil (Youngberg 1958; Dickson et al. 1960; McConnell and Klages 1969; South and Davey 1983; Marx et al. 1984). Since the correlation between Ca and sand content ($r = -0.76$) is negative (South and Davey 1983) sandy soils usually contain <150 $\mu\text{g g}^{-1}$ Ca (Table 2). Some fine-textured soils may contain more than 400 $\mu\text{g g}^{-1}$ Ca (Dickson et al. 1960; Tanaka et al. 1967; Bueno et al. 2012). In one survey, seedling quality of *Pinus strobus* was lower when nursery soil contained more than 200 $\mu\text{g g}^{-1}$ Ca (Dickson et al. 1960). Typically, sandy topsoil in non-fertilized pine plantations contain less extractable Ca than in nursery topsoils (Figure 9).

Approximately 54% of the soils in 43 pine plantations in the southeastern United States contain more than 100 $\mu\text{g g}^{-1}$ Ca (Mehlich 3) and 100% contain more than 15 $\mu\text{g g}^{-1}$ Ca (NCSFNC 1991). In plantations, good growth occurs when topsoil contains 17 to 25 $\mu\text{g g}^{-1}$ Ca (NCSFNC 1991). Although various row-crops exhibit Ca deficient leaves, soils in North America generally have enough Ca so that deficiency symptoms do not occur in pine plantations. As a result, calcium fertilizers are not applied at time of plantation establishment.

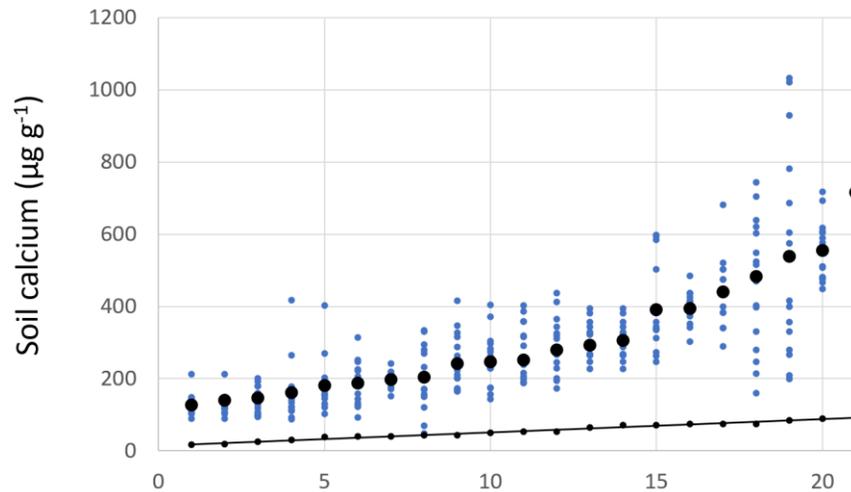


Figure 9. Soil calcium (Mehlich 3) from 21 nurseries and 21 non-fertilized pine plantations in the United States. Fields in nurseries vary from 49 to 1191 $\mu\text{g g}^{-1}$ Ca (blue dots) and soils from the *Pinus taeda* plantations (black line) vary from 17 to 92 $\mu\text{g g}^{-1}$ Ca (NCSFNC 1991). Each nursery is represented by a mean (black dot) of up to 15 soil samples. Calcium fertilizer was applied to nursery fields in March if the soil test was $<200 \mu\text{g g}^{-1}$ (Mehlich 3). In comparison, Ca is typically not applied to pine plantations when extractable soil Ca is below $20 \mu\text{g g}^{-1}$.

5.1 Soil pH

In nursery soil, Ca is positively related to pH mainly because lime is used to increase soil pH (Carter 1987; South and Davey 1983; South et al. 2017). In contrast, most pine plantations are not fertilized with lime and exchangeable Ca is not related to soil pH (NCSFNC 1991; Davis et al. 2007b).

5.2 Organic matter

Organic matter is positively correlated to soil Ca ($r = +0.286$; $p = 0.059$) in fertilized nursery soils (South and Davey 1983). Wilde (1958, p. 364) said that soils with low Ca may be “safely corrected by the addition of organic remains high in bases.” For example, adding leaves (2 cm depth) at the Vallonia Nursery (IN) initially increased soil Ca by $483 \mu\text{g g}^{-1}$ Mg (Davis et al. 2007a). Hardwood leaves may contain $22,700 \mu\text{g g}^{-1}$ Ca and pine bark and sawdust may contain $4,800$ and $800 \mu\text{g g}^{-1}$ Ca, respectively (Mexal and Fisher 1987). Therefore, although more than $44,000 \text{ kg ha}^{-1}$ of sawdust can be applied to seedbeds, available Ca levels might increase by $<20 \mu\text{g g}^{-1}$ Ca (Munson 1982). Adding lime-amended horse manure at the Syracuse Nursery (NY) increased soil to pH 7.3, increased soil Ca to $1,300 \mu\text{g g}^{-1}$, and reduced seedling quality (Bickelhaupt 1989).

5.3 Nitrogen

When nursery seedbeds are fertilized with ammonium fertilizers, Ca levels decrease due to leaching (Wilde and Kopitke 1940; Steinbeck 1962; Knight 1981; Boxman et al. 1991; Dobrahner et al. 2004; Bryson and Mills 2014). Fertilization with 224 kg ha^{-1} N (as AN) reduced soil Ca in Georgia by 17 to $38 \mu\text{g g}^{-1}$ Ca (Figure 10) and in Virginia Ca was reduced by $145 \mu\text{g g}^{-1}$ (Figure 11).

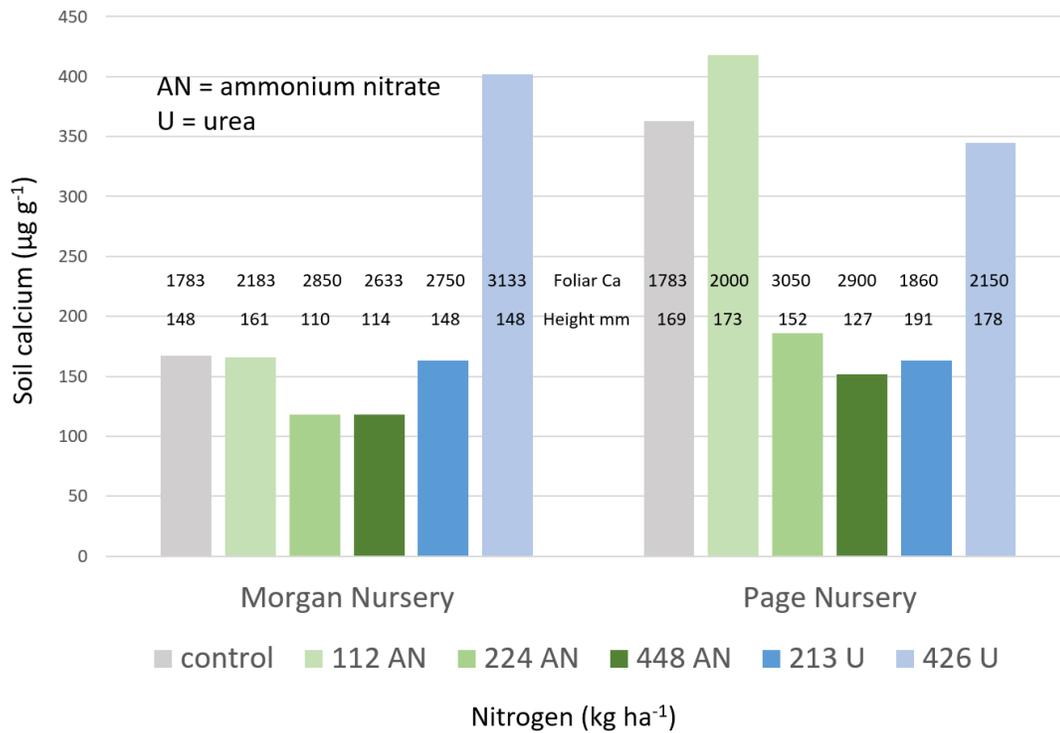


Figure 10. Fertilization of *Pinus elliottii* with ammonium nitrate (AN) increased foliar Ca levels and higher rates reduced the amount of extractable soil Ca (AA) when soil was sampled in December (Steinbeck 1962). This produced a negative relationship between soil Ca and foliar N ($r = -0.56$; $\alpha = 0.01$). The lowest N treatment (112 kg ha⁻¹ of N; source was ammonium nitrate) and both urea treatments (U = Uramite) were applied before sowing (April 17 - Morgan Nursery; April 12 - Page Nursery). April soil samples indicated 87 and 343 µg g⁻¹ Ca for the Morgan and Page nurseries, respectively. High rates of ammonium nitrate were applied using four top-dressings during the month of July. *Pinus elliottii* seedlings at lifting ranged in height from 110 mm to 191 mm and foliar Ca ranged from 1,783 µg g⁻¹ to 3,133 µg g⁻¹. Urea fertilization resulted in “summer chlorosis” on 30 to 70 percent of seedlings from May through August. Fertilization with AN in July resulted in burning of foliage and increased seedling mortality (50% mortality at the Morgan Nursery and 30% at the Page Nursery). More than 70% of these seedlings remained chlorotic until cooler temperatures in October.

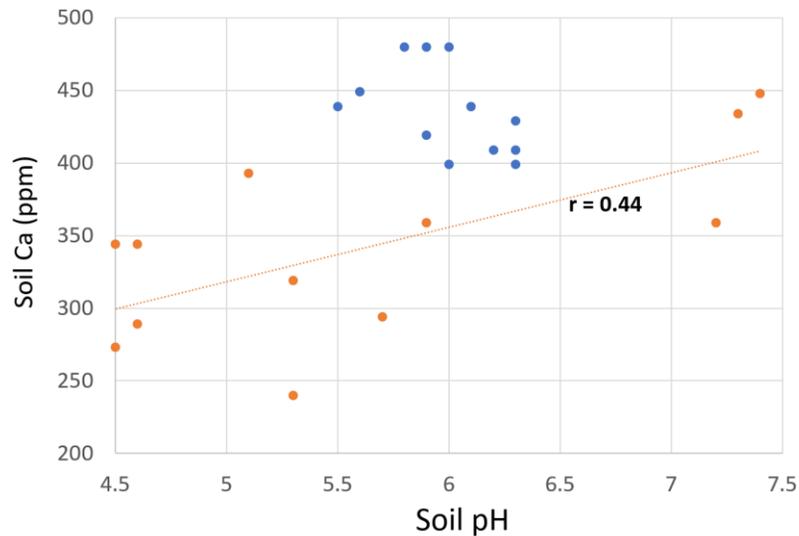


Figure 11. A trial at the Union Camp Nursery (VA) included four sources of N applied at 224, 336 and 448 kg ha⁻¹ N during the growing season (Villarrubia 1980). On June 30, 1978, untreated plots (blue dots) averaged 436 $\mu\text{g g}^{-1}$ Ca (Mehlich 1) and on September 12, 1978, N treated soil (orange dots) averaged 341 $\mu\text{g g}^{-1}$ Ca. Soil treated with ammonium nitrate, ammonium sulfate, urea, and sodium nitrate averaged 279, 320, 357, and 414 $\mu\text{g g}^{-1}$ Ca, respectively. Irrigation and rainfall leached some Ca and some Ca was taken up by seedlings. Fertilization with sodium nitrate reduced soil acidity to pH >7.1, while ammonium sulfate increased acidity to pH <4.7. Urea and ammonium nitrate treatments averaged pH 5.4 and pH 5.2, respectively.

5.4 Phosphorus

Adding CaCO_3 to a pH 5.5 soil increased available P in the soil but decreased foliar P concentrations in *Pinus sylvestris* seedlings to deficiency levels (Carter 1987). In contrast, adding Ca(OH)_2 to a pH 4.8 soil reduced Pt mycorrhiza but did not reduce foliar P of *Pinus taeda* (Marx 1990). If a mycorrhiza-lime interaction exists, this might explain why liming reduces foliar P concentration at some locations but not others. A positive growth response to P fertilization is unlikely when soil Ca is greater than 110 $\mu\text{g g}^{-1}$ Ca (Wells et al. 1973). Some nursery soils exceed 2,000 $\mu\text{g g}^{-1}$ Ca (Martian 1989; Bueno et al. 2012).

A foliar application of H_3PO_4 increased growth of *Pinus taeda* seedlings and reduced foliar Ca concentration by 500 $\mu\text{g g}^{-1}$ (South et al. 1988). A similar response was observed in a greenhouse trial where sodium phosphate increased shoot mass to 23 g (Figure 12). At the Mt. Sopris Nursery (CO), stunted pine seedlings had P-deficient symptoms and foliage contained 1,000 $\mu\text{g g}^{-1}$ P plus 11,300 $\mu\text{g g}^{-1}$ Ca (Landis 1979). Readily available P can combine with Ca to produce an insoluble precipitate.

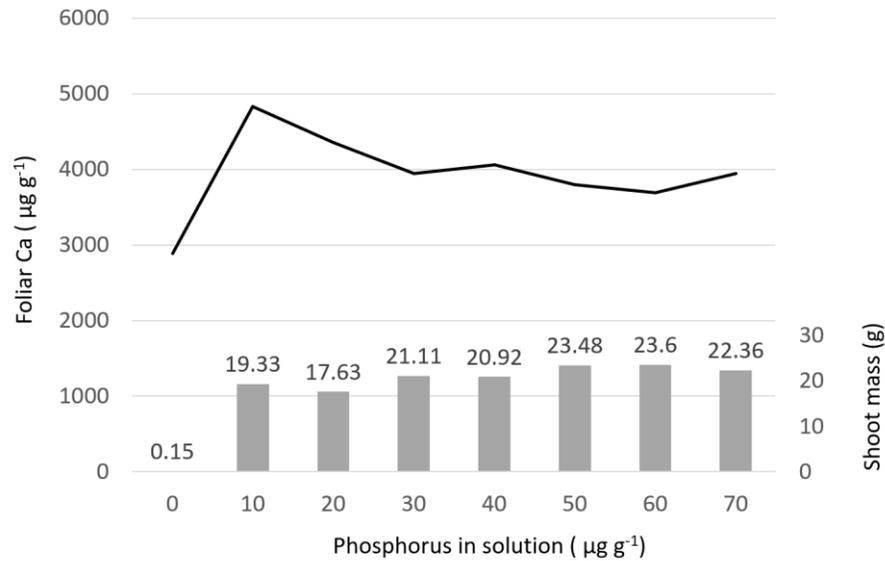


Figure 12. Shoot mass (bars and numbers) and calcium (Ca) in *Pinus taeda* needles (solid line) can be affected by the amount of sodium phosphate in nutrient solutions (Blackmon 1969). Each phosphorus solution also contained $100 \mu\text{g g}^{-1}$ of nitrogen and $100 \mu\text{g g}^{-1}$ of Ca.

5.5 Magnesium

Due to applications of dolomite, Mg and Ca are highly correlated in nursery soil (South and Davey 1983; South et al. 2018). At some nurseries, fertilizing with gypsum before sowing will lower soil Mg levels and induce a temporary Mg-deficiency in pine seedlings (South 2022a). At the Edwards Nursery (NC), fertilizing with CaCO_3 increased soil pH and did not lower soil Mg (Figure 13).

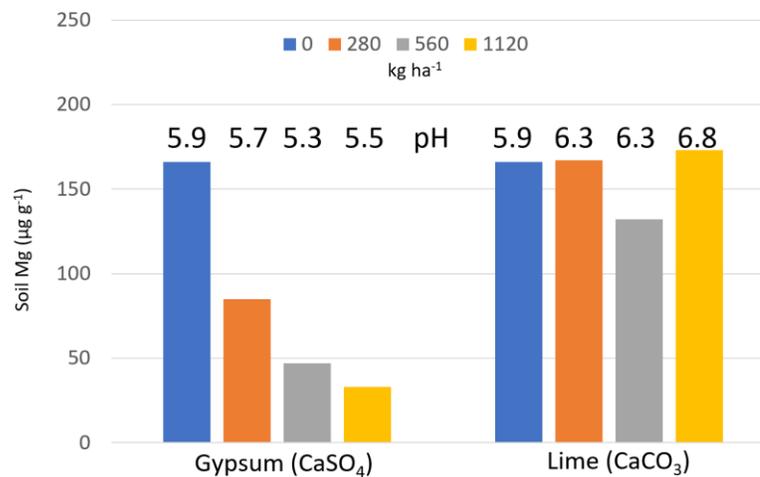


Figure 13. Adding $1,120 \text{ kg ha}^{-1}$ gypsum (29% calcium) to a sandy soil lowered soil pH and reduced available Mg while adding $1,120 \text{ kg ha}^{-1}$ lime (40% calcium) increased soil pH. Treatments at the Edwards Nursery (NC) (Deines 1973) were applied May 18, 1972 and soil was sampled in December 1972. [$\text{pH LSD}_{05} = 0.33$; $\text{Mg LSD}_{05} = 32 \mu\text{g g}^{-1}$].

5.6 Ca/Mg ratio

Soil nutrient ratios in this paper are determined using parts per million ($\mu\text{g g}^{-1}$); not milliequivalents. Those who prefer milliequivalents ratios may divide the Ca/Mg ratios presented here by 1.65.

Apparently, good bareroot pine seedlings were produced using soil Ca/Mg ratios of 12 (McConnell and Klages 1969; Wall 1994), 23 (Rowan 1971) and some “good” seedlings were produced with ratios >40 (Dickson et al. 1960; Landis 1988). In contrast, stunted bareroot pine seedlings occurred in New Zealand with a Ca/Mg ratio of 4 (Will 1961). At the New Zealand nursery, chlorosis had more to do with the absolute amount of Mg ($12 \mu\text{g g}^{-1}$ Mg) and had little to do with a hypothetical imbalance between Ca and Mg. Even so, some believe the balance between Ca and Mg is more “important” than the absolute amount of Mg in the soil [Note: use of the word “important” is subjective without a mathematical basis]. Some say once the Ca/Mg ratio in soil exceeds 10, then managers must add Mg to prevent a reduction in chlorophyll production. While this might be true at some nurseries, there are several examples where ratios were greater than 10 and green seedlings were grown without Mg fertilization (Wall 1994; South 2022a).

Some have questioned the science behind a “Ca/Mg balance” for over a century (Lipman 1916) and data exist to cast doubt on the importance of a low Ca/Mg ratio. Although a soil with a Ca/Mg ratio of 11 can certainly produce yellow-tip needles, it is false logic to assume that all soils with a Ca/Mg ratio above 10 will produce Mg-deficient pine seedlings. Even Wilde questioned the Ca/Mg ratios proposed by Moser (1933). Wilde (1946; p. 83) said no concrete observations verifying such an assumption have thus far been reported in relation to tree growth. Nursery managers successfully grew pines in soil with Ca/Mg ratios as high as 67 (South 2022a). Several agronomists contend the “ideal” nutrient balance theory is flawed (Schulte and Kelling 1985; Kopittke and Menzies 2007; Gaspar and Laboski 2016; Chaganti et al. 2021).

5.7 Manganese

Translocation of Ca into the shoot apex is inhibited by high levels of soil Mn (Marschner 2012) and high soil Mn can stunt pine seedlings (South 2022b). Too much Mn may result in terminal death and resin exudation (i.e. Ca-deficiency) even though samples showed needles contained more than $2,000 \mu\text{g g}^{-1}$ Ca. In a greenhouse trial with saturated soil, applying 81 kg ha^{-1} Mn killed *Pinus resinosa* seedlings (Slaton and Iyer 1974). In one container trial, high levels of Mn and Zn reduced pine roots by 97% (Beyer et al. 2013). The Ca concentration in stunted roots was $<70 \mu\text{g g}^{-1}$ and the concentration in needles was $280 \mu\text{g g}^{-1}$ (Figure 14). Adding dolomite to the soil can increase soil pH and will reduce the risk of a Mn toxicity (South 2022b; Figure 15).



Figure 14. High levels of manganese and zinc in soil reduced shoot and root growth and killed *Pinus strobus* seedlings (Beyer et al. 2013). In this soil (pH 4.1 – 3,200 $\mu\text{g g}^{-1}$ Mn), pine needles had 1,800 $\mu\text{g g}^{-1}$ Mn, 1,200 $\mu\text{g g}^{-1}$ Zn and 280 $\mu\text{g g}^{-1}$ Ca. Leaf mass was 14% of seedlings growing in uncontaminated media. Photo by Nelson Beyer, United States Geological Survey 2009.

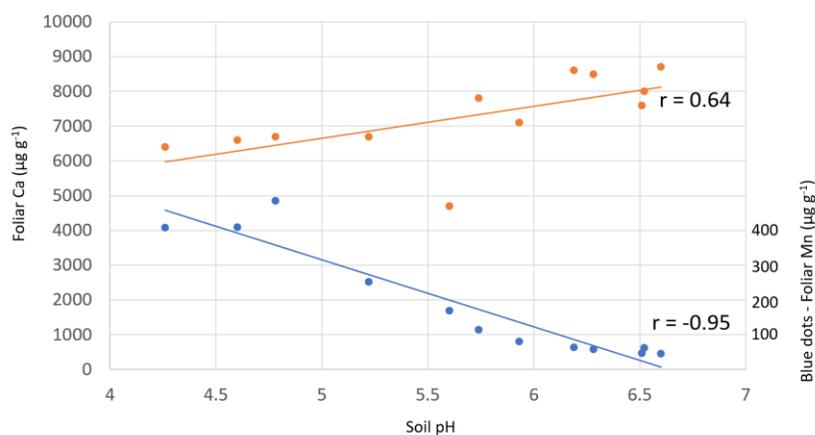


Figure 15. Adding dolomite to potting media increased soil pH, increased foliar Ca (orange dots) and reduced the uptake of manganese (Mn) in *Pinus rigida* foliage (Helm and Kuser 1991). Soil at pH 4.26 had no added lime while pH 6.6 soil was achieved by adding 40 g of dolomite per kg of soil. Seedling growth was maximum at pH 4.8.

6 Irrigation water

Ca in irrigation water at many nurseries is enough to meet the needs of pine seedlings (Carlson 1979; Landis 1979; Argo et al. 1997; Landis et al. 2009). When pines were growing in sand with low available Ca (14 $\mu\text{g g}^{-1}$) and were irrigated with water containing 8 mg L^{-1} of Ca, the needle concentration exceeded 4,000 $\mu\text{g g}^{-1}$ Ca (Steinbeck 1962). About 93% of irrigation water samples from southern nurseries contain more Ca than 8 mg L^{-1} (Figure 16). Applying 600 mm of irrigation (at 10 mg L^{-1} Ca) is equivalent to applying 286 kg ha^{-1} of gypsum (21% Ca).

At one sandy location, irrigating non-fertilized soil for 16 years (632 mm year^{-1}) increased available soil Ca by 75 kg ha^{-1} (Albaugh et al. 2014). In addition to irrigation,

1,000 mm of rainfall may provide 3 to 38 kg ha⁻¹ of Ca (Madgwick and Ovington 1959; Carroll 1962; Allen et al. 1968).

When growing plants in greenhouses, a few researchers believe “The calcium and magnesium ratio in the substrate solution (and in the irrigation water) should be 3 Ca to 1 Mg if expressed as meq/L or 5 Ca to 1 Mg if expressed as ppm Ca and Mg” (Bailey et al. 1999). Even so, there are insufficient data to make the Ca/Mg ratio “an evaluation factor when judging the suitability of a water for irrigation” (Ayers and Westcot 1985; South 2022a). At one *Pinus taeda* nursery, irrigation water contained 50 µg L⁻¹ Ca and 2 µg L⁻¹ Mg (McNabb and Heidbreder-Olson 1998). To avoid producing yellow-tip needles, foliar levels at that nursery were checked monthly and, if needed, Mg was applied over the top of seedlings in July.

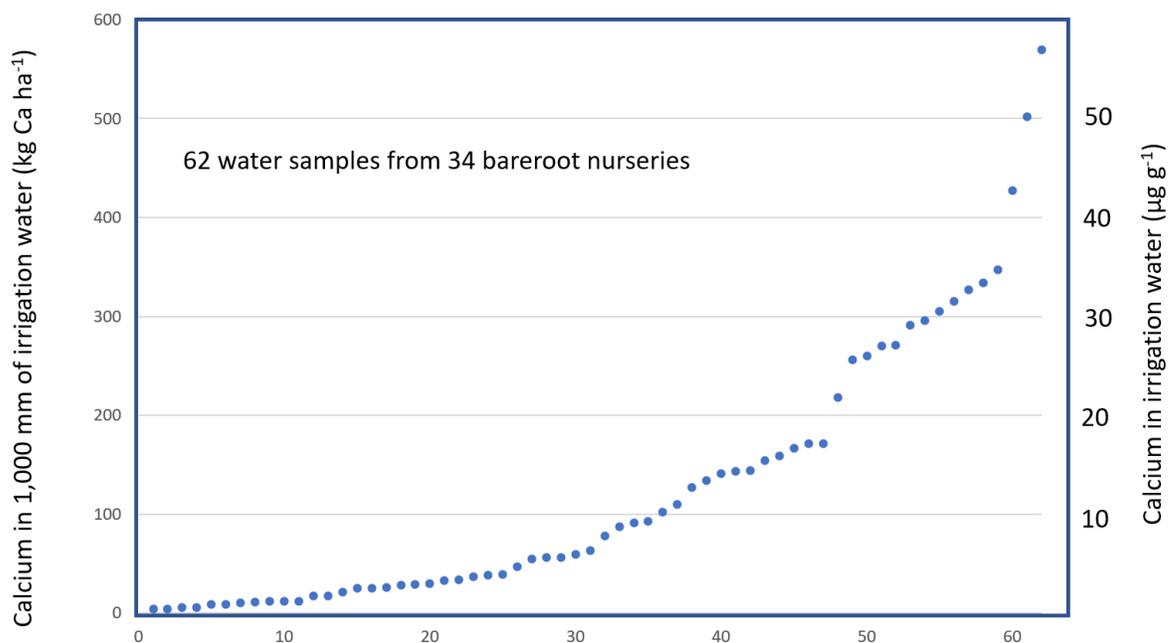


Figure 16. The amount of soluble Ca in irrigation water from 34 nurseries ranged from 4 to 57 mg L⁻¹ (McNabb and Heidbreder-Olson 1998). When the level of Ca in irrigation water is 10 mg L⁻¹, then 60 cm of irrigation would add approximately 60 kg ha⁻¹ year⁻¹. At four nurseries, the Ca/Mg ratio (µg) varied from 11 to 525.

7 Mycorrhiza

Pine seedlings without mycorrhiza do not become Ca-deficient (Ingestad 1962; Bücking et al. 2002). In a bareroot nursery, non-mycorrhizal *Pinus taeda* seedlings exhibited P deficiency symptoms while needles contained >3,000 µg g⁻¹ of Ca (South et al. 1988). Adequate foliar Ca concentrations were also observed for non-mycorrhizal seedlings in greenhouse tests (Cumming and Weinstein 1990; Walker and McLaughlin 1997; Zhang and George 2010).

High levels of soil Ca can reduce the formation of Pt ectomycorrhiza (Marx 1990) and, in one trial, Pt ectomycorrhiza was inversely correlated ($r = -0.36$) with foliar Ca concentration (Mitchell et al. 1990). Likewise, a lack of mycorrhizal infection was noted when container-grown *Pinus sylvestris* seedlings were fertilized with Ca-nitrate (Kieliszewska-Rokicka 1991).

8 Calcium removed at harvest

Depending on species, cultural practices, and seedling age, a million pine 1-0 seedlings may contain 6 to 20 kg of Ca (Schenck 1907; Lunt 1938; Knight 1978b; Flinn et al. 1980; Donald and Young 1982; South and Boyer 1983; Nelson and Switzer 1985; Simpson 1985; Dobrahner et al. 2004). Harvesting 1.7 million pine seedlings might remove 26 kg ha⁻¹ of Ca while harvesting *Zea mays* grain removes about a tenth of that amount (Heckman et al. 2003).

The Ca levels in topsoil decline over time when harvest rates exceed inputs from irrigation, fertilizers, and rainfall (Will and Knight 1968). Field #7, at the Ashe Nursery in Mississippi, had 380 µg g⁻¹ Ca in 1969 and 68 µg g⁻¹ in 1981 which equates to an average decline of 26 µg g⁻¹ year⁻¹. Likewise, at the Lava Nursery (OR) and Duncan Nursery (BC), the decline was 33 µg g⁻¹ year⁻¹ (White et al. 1980; Youngberg 1984). At one area of the Westvaco Nursery (SC), rainfall and N-fertilization of cover-crops resulted in an average annual loss of 21 µg g⁻¹ Ca from 1983 to 1990 (Figure 17).

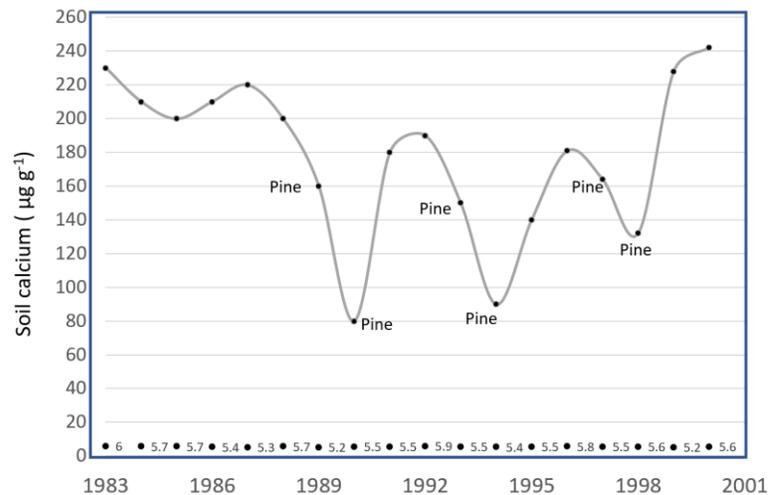


Figure 17. Soil calcium levels (Mehlich 1) at a bareroot nursery in South Carolina. Field B-1 was managed with cover-crops from 1983 to early 1989. *Pinus taeda* seed were sown in April of 1989, 1990, 1993, 1994, 1997 and 1998. Soil pH values adjacent to dots represent soil acidity in October-November. Dolomitic limestone was applied in 1988 (1,120 kg ha⁻¹), 1991 (2,240 kg ha⁻¹), 1992 (1,456 kg ha⁻¹) and 1998 (1,120 kg ha⁻¹).

9 Calcium concentration after transplanting

In general, outplanting performance is not related to the foliar Ca of pine seedlings at time of lifting (Madgwick 1964; Larsen et al. 1988; van den Driessche 1991). After tree planters cover roots with soil, pine seedlings take up enough cations to avoid a Ca-deficiency. In Texas, *Pinus echinata* needles had 724 µg g⁻¹ Ca when outplanted in March and eight months later they contained 1,100 to 2,750 µg g⁻¹ Ca (Bryson 1980). In 1979 and 1980, outplanted *Pinus ponderosa* seedlings ended the growing season at the same foliar concentration as at planting in late March (Baer 1984).

10 Toxicity

When pine needles contain less than $5,000 \mu\text{g g}^{-1}$ Ca (Table 5), seedlings do not show toxicity symptoms. However, stunted seedlings may result when the concentration in foliage exceeds $6,500 \mu\text{g g}^{-1}$ Ca (van Lear and Smith 1972; Carter 1987; Landis 1988; Potvin et al. 2014; Hachani et al. 2020; Figure 4). Occasionally pine foliage exceeds $9,000 \mu\text{g g}^{-1}$ Ca (Table 5). At the Mt. Sopris Nursery (CO), chlorotic, Fe-deficient, pine needles contained $11,300 \mu\text{g g}^{-1}$ Ca (Landis 1979).

Too much carbonates in soil can reduce height growth of pines (Erdmann 1966; Carter 1987) and too much Ca-acetate ($3,735 \mu\text{g g}^{-1}$ Ca) can increase damping-off of *Pinus echinata* seedlings (Chapman 1941). Too much lime can cause deficiencies in Fe, B (Stone et al. 1982; Shorrocks 1997; South 2021) and Mn (Kishchuk 2000). Occasionally, too much gypsum will cause a temporary Mg deficiency in acid soils. When S is applied just before sowing, gypsum crystals may form on roots and this could reduce growth of pine seedlings (Carey et al. 2002). When soil contains more than $1,200 \mu\text{g g}^{-1}$ Ca, a B-deficiency may emerge (Gupta and MacLeod 1981; Stone et al. 1982; Landis 1988; South 2021). The toxicity threshold for Ca in irrigation water is $1,000 \mu\text{g g}^{-1}$ Ca (Landis et al. 1989).

High levels of Cl are toxic to pine seedlings, and therefore too much CaCl_2 can kill or stunt seedlings growing in sand (Mitchell 1939; Pharis et al. 1964; Timmer and Parton 1984). Chlorosis increased when a *Pinus resinosa* plantation was fertilized with 342 kg ha^{-1} CaCl_2 (Stone 1953). High rates of $\text{Ca}(\text{OH})_2$ can increase soil pH and reduce growth of pine (Marx 1990). Most pines are not tolerant of soil with high salt levels and/or high soil pH and these soils often contain high levels of Ca (Carter 1987; Landis 1988; Hachani et al. 2020). Some say naturally high pH soils should never be selected for a forest nursery (Aldhous 1975; Landis 1988).

11 Ca Fertilizers

11.1 CaCO_3

When applied to a nursery soil at $1,100 \text{ kg ha}^{-1}$, agricultural lime (CaCO_3) increased a nursery soil pH by almost 1 unit (Figure 18). Although soil laboratories may suggest adding $2,000 \text{ kg ha}^{-1}$ of lime when a nursery soil is at pH 5.0, this rate is not recommended for pine nurseries. In some cases, applying more than $1,500 \text{ kg ha}^{-1}$ of lime can reduce growth of pine in greenhouses (Table 8).

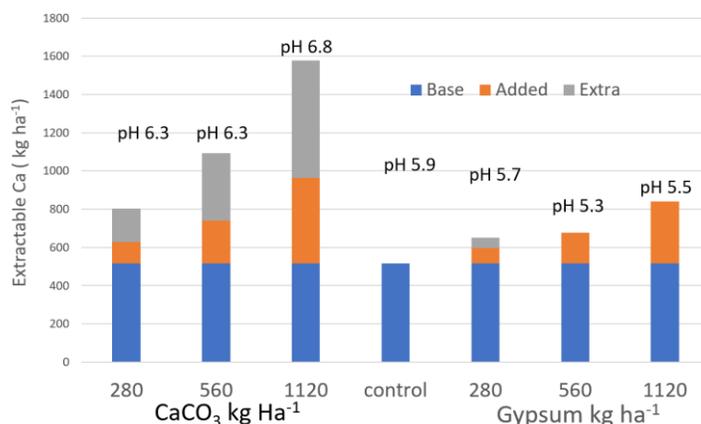


Figure 18. Fertilization (May 18, 1972) with calcium carbonate (agricultural lime – 40% Ca) and calcium sulfate (gypsum – 29% Ca) increased the amount of calcium in a silt loam soil in November at the Edwards Nursery in North Carolina (Deines 1973). Solubility may explain why soil Ca in November was lower than expected for gypsum treatments with pH<5.6. Solubility of calcium carbonate in water is 13 mg per liter and gypsum solubility may range from 2,000 to 2,500 mg per liter. Extra Ca in November (greater than amount applied) likely represents cations retained due to pH>6.0. LSD₀₅ = 330 kg ha⁻¹

Table 8. The effect of CaCO₃ fertilization on heights (mm) and mass (g) of pines grown outside in soil (McIntyre and White 1930; Wahlenberg 1930; Lunt 1947) or in greenhouses. Recommended soil acidity for pine seedbeds is pH 4.5 to 5.5. pH = before treatment.

Species	pH	Rate	Control	CaCO ₃	change	Reference
		kg ha ⁻¹	-mm-	-mm-		
<i>Pinus ponderosa</i>	5.3	3,025	64	52	-19%	Wahlenberg 1930
<i>Pinus radiata</i>	6.3	40,000	64	57	-12%	Nakos 1979
<i>Pinus strobus</i>	--	2,240	55	47	-14%	McIntyre and White 1930
<i>Pinus sylvestris</i>	5.5	4,000	64	50	-22%	Carter 1987
<i>Pinus taeda</i>	3.0	2,337	96	78	-19%	Coultas et al. 1991
			-g-	-g-		
<i>Pinus caribaea</i>	5.8	1,792	8.09	4.68	- 42%	Richards and Wilson 1963
<i>Pinus caribaea</i>	6.3	1,792	7.18	7.61	+6	Richards and Wilson 1963
<i>Pinus elliotii</i>	4.3	2,694	13.5	13.2	-2%	McKee 1978
<i>Pinus resinosa</i>	5.0	8,960	40.9	35.5	-13%	Lunt 1947
<i>Pinus strobus</i>	5.6	2,240	90.0	108.6*	+20%	McIntyre and White 1930
<i>Pinus strobus</i>	--	2,240	31.1	40.4*	+30%	McIntyre and White 1930
<i>Pinus sylvestris</i>	4.6	6,000	0.107	0.072	-33%	Wallander et al. 1997
<i>Pinus taeda</i>	6.7	3,136	2.18	0.67	- 69%	Richards 1965
<i>Pinus taeda</i>	3.5	2,240	14.1	13.1	- 7%	Barbour and Berenyl 1969

* Damping-off might have increased seedling mass

A target value of 200 µg g⁻¹ Ca for bareroot pine seedbeds (South and Davey 1983) is supported by *Pinus strobus* data (Dickson et al. 1960) and by a greenhouse test with *Pinus elliotii* (Figure 19). In contrast, targets of 300, 500 or 1,250 µg g⁻¹ Ca are based on assumptions. As a result, Ca applied to meet these targets could be a waste of time and money (Crowther 1950; Dickson et al. 1960; McKee 1978; Bickelhaupt 1989).

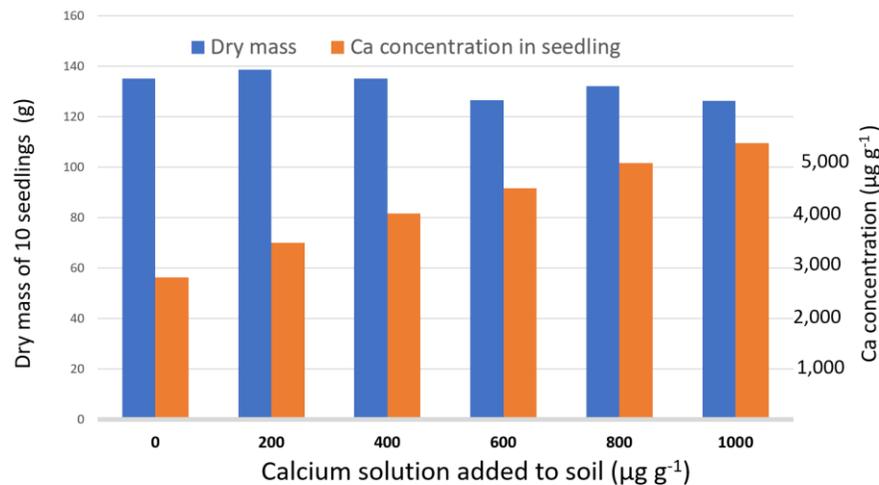


Figure 19. The effect of CaCO_3 on growth (dry mass) and foliar concentration of *Pinus elliottii* seedlings in a greenhouse (McKee 1978). Each pot contained 10 seedlings growing in a Caddo silt loam and untreated soil contained $318 \mu\text{g g}^{-1}$ Ca. Pots treated with $400 \mu\text{g g}^{-1}$ Ca (equivalent in this trial to 422 kg ha^{-1}), increased Ca concentration (total seedling) but did not improve seedling growth. It seems that increasing Ca concentration in foliage to more than $4,000 \mu\text{g g}^{-1}$ decreased growth ($Y = 148 \text{ g} - 0.0038 * \mu\text{g g}^{-1} \text{ Ca in seedling}$; $n = 6$; $r = -0.75$). Soil averaged pH 4.3 at the start of the study and, after 51 weeks, soil with the highest rate of CaCO_3 was at pH 5.8.

11.2 $\text{Ca}(\text{OH})_2$

Calcium hydroxide is typically not used by nursery managers but researchers sometimes apply it because it does not contain Mg, P or S (Crannell et al. 1994). In one trial, 2.85 Mg ha^{-1} of $\text{Ca}(\text{OH})_2$ applied before sowing reduced growth of *Pinus taeda* (family 1-68) seedlings (Marx 1990). The treatment increased soil pH to 5.8 and seedling biomass was reduced by 21% (seedlings were also treated with 150 kg ha^{-1} of N). The increase in soil pH also reduced growth of *Pt mycorrhiza*.

11.3 $\text{CaCO}_3 \cdot \text{MgCO}_3$

Soil and dolomite represent the primary sources of Ca in bareroot nurseries. The amount of dolomite applied varies with soil pH, CEC and year. Applying too much dolomite can increase Fe chlorosis (Shoulders and Czabator 1965) and damping-off (Wilde 1942; South 2017). At the Westvaco Nursery, dolomite was applied three times from 1988 to 1998 (Figure 16) which equates to 81 kg year^{-1} of Ca. Although contents vary with region and distributor, 1,000 kg of dolomite usually contains 220 kg of Ca.

11.4 $\text{CaSO}_4 \cdot 2\text{H}_2\text{O}$

Sometimes gypsum might initially decrease soil pH by 0.4 unit (Fried and Peech 1946; Figure 18) but in other soils pH might increase by 0.4 unit (Marx 1990). No effect on pH is expected when gypsum is applied to nursery soils above pH 4.5-5.0. Although some researchers apply gypsum at $>1,700 \text{ kg ha}^{-1}$, nursery managers are reluctant to apply more than 800 kg ha^{-1} since higher rates might result in a Mg deficiency in sandy soils (South 2022a). Several nursery trials with gypsum have failed to demonstrate an

increase in pine biomass (Table 10). Since gypsum is soluble, Ca does not last very long in sandy topsoils (Figure 18).

Applying gypsum before sowing will reduce the risk of a S deficiency. A rate of 870 kg ha⁻¹ would provide 200 kg ha⁻¹ S (Table 1). A S-deficiency might partly explain why lowering soil pH with H₂SO₄ increased growth of *Pinus strobus* seedlings (Wood and Bormann 1977). S-deficiencies have been reported at a few conifer nurseries (Lyle and Pearce 1968; Bolton and Benzian 1970; Morris 1979).

Some managers notice yellow-tip needles on pine seedlings growing in fields fertilized with N and gypsum (CaSO₄•2H₂O). To reduce the risk of a Mg-deficiency at sandy nurseries, the rate of gypsum can be lowered (e.g. <800 kg ha⁻¹). However, the risk of yellow-tip symptoms is low at nurseries with CEC >50 meq kg⁻¹. At some nurseries, applying 1,500 kg ha⁻¹ of gypsum (Table 9) did not produce a Mg deficiency.

It seems doubtful that applying gypsum to nursery soil will increase freeze tolerance of conifers. For example, applying gypsum to potted saplings did not increase freeze tolerance of *Picea rubens* (Schaberg et al. 2000) and applying extra Ca to *Pinus elliotii* did not increase cell wall thickness (Malavolta et al. 1970).

Table 9. The effect of calcium sulfate (gypsum) fertilization on heights (mm) or mass (g) of pines in bareroot nurseries.

Species	Rate kg ha ⁻¹	Control -mm-	Gypsum -mm-	change	pH	Reference
<i>Pinus ponderosa</i>	4,116	64	70	+10	5.3	Wahlenberg 1930
<i>Pinus ponderosa</i>	8,232	64	70	+12	5.3	Wahlenberg 1930
<i>Pinus radiata</i>	140	188	188	0%	5.0	Will 1961
<i>Pinus taeda</i>	6,776	278	238	-14%	5.4	South 2021
		-g-	-g-			
<i>Pinus palustris</i>	1,568	13.5	11.2	-17%	--	Maki and Henry 1951
<i>Pinus palustris</i>	1,568	12.7	13.6	+7%	--	Maki and Henry 1951
<i>Pinus radiata</i>	1,500	2.62	2.34	-11%	5.0	Flinn and Waugh 1983
<i>Pinus taeda</i>	5,300	9.4	9.7	+3%	5.1	Marx 1990
<i>Pinus taeda</i>	10,600	9.4	9.7	+3%	5.3	Marx 1990

11.5 Ca(NO₃)₂

As with most fertilizers, application rate is important. In one container trial, applying 2,665 kg ha⁻¹ of Ca-nitrate reduced average height of 18-week-old *Pinus sylvestris* (Holopainen et al. 1995). Likewise, dipping needles in a solution containing 4,000 ppm N produced slight burning of needle tips (Eberhardt and Pritchett 1971). Nursery managers, however, routinely apply Ca-nitrate without causing foliar injury (Dumroese and Wenny 1997). At one bareroot nursery with high soil Ca, the annual amount of Ca-nitrate applied to *Pinus strobus* totaled 1,160 kg ha⁻¹; 180 kg ha⁻¹ N and 220 kg ha⁻¹ Ca (Dobrahner et al. 2004). The product was sprayed over the top of pine seedlings at a rate of 22 kg ha⁻¹ of Ca. In 1982, a similar rate was applied over *Pinus taeda* seedlings at the Ft. Towson Nursery (OK).

In greenhouse trials using soil, pines fertilized with Ca-nitrate grew the same or better than those fertilized with ammonium nitrate or ammonium sulfate (Addoms 1937; Pharis et al. 1964; Kieliszewska-Rokicka 1991). Some say Ca-nitrate is the proper source when applying N in the autumn (Landis 1996). Even so, some believe nitrate is an inefficient source of N (Weetman and Algar 1974; Davey 1988).

Since Ca-nitrate has 15.5% N compared to 32% N for 32-0-0, most bareroot managers use economics to decide which form of N to purchase. When both products cost the same per tonne, most bareroot managers will choose 32-0-0 since a dollar will purchase twice the amount of N as Ca-nitrate.

Applications of Ca-nitrate increased freeze tolerance of deciduous trees (Percival and Barnes 2008) and applications of ammonium nitrate increased freeze tolerance of *Pinus palustris* and *Pinus halepensis* (Davis et al. 2011; Toca et al. 2018). In contrast, applying CaCl_2 did not increase freeze tolerance of *Pinus sylvestris* (Christersson 1973, 1975). It follows that if Ca-nitrate increases freeze tolerance, it likely is due to added N.

11.6 CAN

Calcium ammonium nitrate (CAN) exists in several forms. Some contain limestone, some contain dolomite and others contain calcium carbonate (Maxwell 2012). When calcium carbonate is added to AN, a granular product (26% or 27% N) is sold as “Nitro-Chalk” in the United Kingdom (Benzian 1959; Atterson 1969; O'Reilly et al. 2008), CAN in New Zealand (Ballard and Will 1978) and Cal-am in Australia (May et al. 2009). Adding calcium carbonate reduces the sensitivity of ammonium nitrate to detonation. The granular product (27-0-0) has been applied at rates up to 600 kg ha^{-1} (O'Reilly et al. 2008). A liquid product (17-0-0), made by adding Ca-nitrate to AN, is used to grow seedlings in containers (Dumroese and Wenny 1997). Since soil typically contains sufficient Ca, bareroot nursery managers tend to favor N fertilizers that lower soil pH but do not contain Ca. In contrast, use of CAN has little effect on soil acidity. In 1975, CAN represented about 6% of the N fertilizers used in New Zealand forestry (Ballard and Will 1978).

11.7 CaCl_2

An application of 22 kg ha^{-1} Cl is applied to certain row-crops when topsoil contains $<15 \mu\text{g g}^{-1}$ Cl (Diaz 2019) but low Cl levels is not a concern in tree nurseries (Donald 1991). In greenhouses, CaCl_2 has been applied to pines 5 weeks after sowing (Timmer and Parton 1984; Dumroese and Wenny 1997) but a few managers prefer CAN which contains no Cl_2^- . Applying too much Cl_2^- can damage or kill pine seedlings in greenhouses (Sucoff 1962; Pharis et al. 1964; Heidmann and Thorud 1976; Sands and Clarke 1977). Applying CaCl_2 can increase freeze tolerance of *Pyrus communis* (Rease 1996) but does not increase freeze tolerance of pine (Pellett and Carter 1981).

Pinus taeda seedlings were affected by too much Cl_2^- when they were grown in pots containing white sand for 4.5 months (Pharis et al. 1964). Ten fertilizer treatments had 100% survival but seedlings treated with a double dose of Cl_2^- (ammonium chloride plus calcium chloride) had 46% mortality (11/24). Likewise, 17% died (4/24) when treated with a high rate of ammonium chloride.

In a greenhouse trial with *Pinus virginiana*, seedlings were fertilized with 5 rates of Ca and 3 rates of Mg (Sucoff 1962). The best growth occurred with $20 \mu\text{g g}^{-1}$ Ca and $24 \mu\text{g g}^{-1}$ Mg (Figure 20).

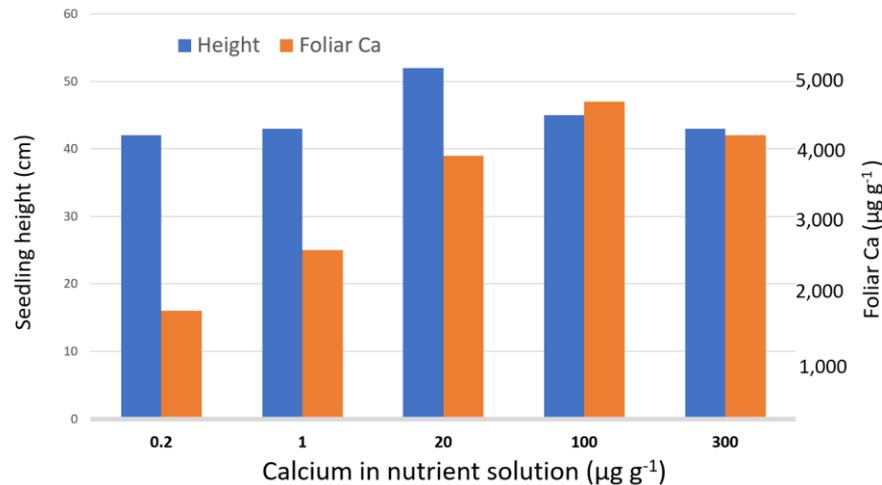


Figure 20. The effect of calcium chloride on height growth of *Pinus virginiana* growing in sand when magnesium is solution is $24 \mu\text{g g}^{-1}$ (Sucoff 1962). There was no significant difference in seedling height ($\alpha = 0.05$). The two highest Ca solution rates contained $278 \mu\text{g g}^{-1}$ Cl and $833 \mu\text{g g}^{-1}$ Cl.

11.8 $\text{Ca}(\text{H}_2\text{PO}_4)_2 \cdot \text{H}_2\text{O}$

In 1992, more than 34,400 kg of TSP were applied to nursery soils in the southern United States which was equivalent to 10 kg Ca per million seedlings (South and Zwolinski 1996). Prior to sowing, the IFA Nursery (WA) applied 224 kg ha^{-1} of TSP which supplied 33.6 kg ha^{-1} Ca (Marx et al. 1984). More than 500 kg ha^{-1} of TSP was applied to fallow fields at the Big Sioux Nursery (SD) (Martian 1989). The Bend Nursery (OR) and Griffith Nursery (NC) also applied TSP over the top of seedlings (Gleason 1989; Hinesley and Maki 1980).

Application of TSP increased available P to more than $50 \mu\text{g g}^{-1}$ P (Mehlich 3) at several nurseries. Although TSP use in North America has declined, it is still used to fertilize pine plantations in some countries (May et al. 2009).

11.9 $\text{Ca}(\text{H}_2\text{PO}_4)_2 \cdot \text{CaSO}_4$

In 1977, 224 kg ha^{-1} of ordinary super phosphate (OSP) was applied before sowing at nurseries in Florida and Louisiana (Marx et al. 1984). This rate supplied 43 kg ha^{-1} Ca, 18 kg ha^{-1} P and 25 kg ha^{-1} S. When managers stopped using OSP, a S deficiency occurred at several nurseries. As a result, use of gypsum in bareroot nurseries has increased since 1960.

When applied before sowing at two nurseries in Georgia, OSP increased height growth at one nursery (Figure 21). In contrast, applying OSP before sowing did not improve growth at two *Pinus taeda* nurseries (Rosendahl and Korstian 1945; Switzer and Nelson 1956). Due to a low P content, OSP is rarely used at bareroot nurseries in North America but it is used in Africa, Asia and South America.

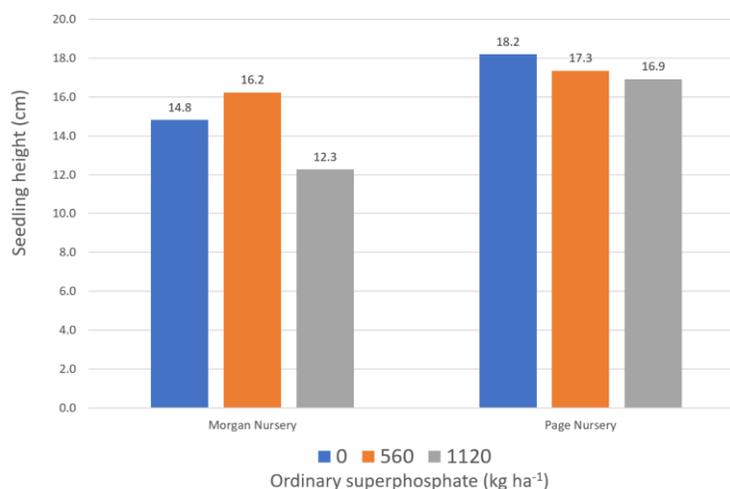


Figure 21. Fertilization (March-April, 1961 prior to sowing) with ordinary superphosphate (560 kg ha⁻¹) increased height growth of *Pinus Elliottii* at the Morgan Nursery (88 µg g⁻¹ P in soil) and had no effect at Page Nursery (129 µg g⁻¹ P in soil) in Georgia (Steinbeck 1962). The 1,120 kg ha⁻¹ rate added approximately 224 kg ha⁻¹ Ca and 123 kg ha⁻¹ S.

11.10 Chelates

Researchers may test Ca-chelates when row-crops show deficiency symptoms. However, a lack of deficiency symptoms in pine nurseries explains why few if any chelate experiments have been installed in bareroot nurseries. Most managers see no reason to apply Ca-chelates to bareroot pines. Pine needles have a thick epicuticular wax layer. In addition, due to near immobility of Ca, foliar applications are likely not adequate to correct a deficiency (Prado 2021).

11.11 Unknown anions

Several researchers published Ca experiments without including the associated anion (e.g. Cl⁻, NO₃, OH). Some likely believed Cl is an inert element and they saw no need to mention it in the methods section. When the rate of Ca-x increased root length of *Pinus massoniana*, was extra growth due to Cl⁻, or NO₃ or Ca? Likewise, when a researcher concludes that CaNO₃ is not a good fertilizer for pine, was it because the researcher cited hydroponic trial with NaNO₃? Invalid conclusions have been published because someone assumed pine growth was not affected by anions.

12 Costs

Fertilizer costs vary by region, shipping distance, year, and distributor. Cost comparisons are time sensitive and vary by region. To keep Ca costs below \$2 kg⁻¹, managers in the United States typically fertilize with dolomitic limestone to increase soil pH and apply gypsum when pH is near optimum. When gypsum costs \$0.26 kg⁻¹, the price of Ca would equal \$1.13 kg⁻¹ (assuming no value for S). When Mg and Ca have equal value, then Ca from dolomite may cost \$0.75 kg⁻¹. In comparison, Ca might cost \$2 kg⁻¹ when applied as Ca-nitrate (assuming N has the same value as N in urea). At nurseries that sell pine seedlings for 7 cents each, managers often purchase low-cost sources of Ca and N fertilizers.

13 Conclusions

- (1) When nursery soil pH is above 5.0, no beneficial effect has been demonstrated by applying CaCO_3 to pine seedbeds.
- (2) Most published critical values for Ca in needles (i.e., low foliar concentration resulting in a 10% growth reduction) were not determined using fertilizer response curves. The assumption that all conifers have foliage with a “greenhouse” critical value of $1,200 \mu\text{g g}^{-1}$ Ca is invalid.
- (3) Due to taking a short-cut method, recommendations of $2,800 \text{ kg ha}^{-1}$ Ca for nursery soils (Wilde 1938) are about $2,000 \text{ kg ha}^{-1}$ too high for *Pinus strobus* seedbeds.
- (4) When irrigation water provides more than 80 kg ha^{-1} Ca during a year, there is no need to apply Ca before sowing pine seed.
- (5) When growing in soil, non-mycorrhizal pine seedlings do not become Ca-deficient.
- (6) There is no proof of a hidden hunger for Ca when *Pinus taeda*, *Pinus elliottii*, *Pinus echinata*, *Pinus virginiana* and *Pinus palustris* are growing in irrigated bareroot nurseries.
- (7) When lime increases damping-off of seedlings, growth of surviving pines may be increased due to a reduction in competition for light, water and nutrients.
- (8) When irrigation water contains less than $3 \mu\text{g g}^{-1}$ Mg, applying too much gypsum before sowing can induce a Mg deficiency in pine seedlings.
- (9) In rare cases when gypsum fertilization increases height growth, the pine seedlings were likely deficient in S.
- (10) When pine needles contain $>900 \mu\text{g g}^{-1}$ Ca, there is no need to apply CaNO_3 to bareroot pine seedlings during the hardening phase in the fall. However, it is not known what might happen when pine needles contain $300 \mu\text{g g}^{-1}$ Ca.
- (11) Chloride is not inert and too much in hydroponics can kill pine seedlings. Some conclusions were invalid because authors assumed pine growth was not affected by anions.
- (12) Fine-textured soils with $>3,000 \mu\text{g g}^{-1}$ extractable Ca should not be selected for a bareroot pine nursery.

14 Acknowledgments

I thank members of the Southern Forest Nursery Management Cooperative for providing soil and foliage data from pine nurseries. I thank John Mexal, J.B. Jett, and Chase Weatherly for feedback on earlier drafts.

15 References

- Addoms RM (1937) Nutritional studies on loblolly pine. *Plant Physiol* 12(1): 199-205. <https://doi.org/10.1104/pp.12.1.199>
- Albaugh TJ, Kiser LC, Fox TR, Allen HL, Rubilar RA, Stape JL (2014) Ecosystem nutrient retention after fertilization of *Pinus taeda*. *Forest Sci* 60(6): 1131-1139. <http://dx.doi.org/10.5849/forsci.13-159>
- Aldhous JR (1975) Nursery Practice. Forestry Commission Bull. 43, London, UK. <https://cdn.forestresearch.gov.uk/1972/03/fcbu043.pdf>
- Allen HL (1987) Forest fertilizers. *J Forest* 85(2): 37-46. <https://doi.org/10.1093/jof/85.2.37>
- Allen SE, Carlisle A, White EJ, Evans CC (1968) The plant nutrient content of rainwater. *The Journal of Ecology* 56(2): 497-504. <https://doi.org/10.2307/2258247>
- Alva AK (1993) Comparison of Mehlich 3, Mehlich 1, ammonium bicarbonate-DTPA, 1.0 M ammonium acetate, and 0.2 M ammonium chloride for extraction of calcium, magnesium, phosphorus, and

- potassium for a wide range of soils. *Commun Soil Sci Plan* 24(7-8): 603-612. <https://doi.org/10.1080/00103629309368826>
- Andrews JA, Johnson JE, Torbert JL, Burger JA, Kelting DL (1998) Minesoil and site properties associated with early height growth of eastern white pine. *J Environ Qual* 27(1): 192-199. <https://doi.org/10.2134/jeq1998.00472425002700010027x>
- Armson KA, Sadreika V (1979) Forest tree nursery soil management and related practices. Ontario Ministry of National Resources. 179 p. https://www.google.com/books/edition/Forest_Tree_Nursery_Soil_Management_and/f3oqAQ_AAMAAJ?hl=en
- Arnold MA, Struve DK (1993) Root distribution and mineral uptake of coarse-rooted trees grown in cupric hydroxide-treated containers. *HortSci* 28(10): 988-992. <https://doi.org/10.21273/HORTSCI.28.10.988>
- Argetsinger LM (1941) Chemical fertilizer treatments of Norway pine transplants in University of Michigan nursery. MF thesis. University of Michigan, Ann Arbor. 64 p. <https://deepblue.lib.umich.edu/bitstream/handle/2027.42/113806/39015003283861.pdf?sequence=1>
- Argo WR, Biernbaum JA, Warncke DD (1997) Geographical characterization of greenhouse irrigation water. *HortTechnology* 7(1): 49-55. <https://doi.org/10.21273/HORTTECH.7.1.49>
- Atterson J (1969) Fertiliser research in Scottish nurseries and forests. *J Sci Food Agr* 20(7): 415-416. <https://doi.org/10.1002/jsfa.2740200711>
- Auten JT (1945) Response of shortleaf and pitch pines to soil amendments and fertilizers in newly established nurseries in the central states. *J Agric Res* 70(12): 405-426. <https://babel.hathitrust.org/cgi/pt?id=uva.x004385460&view=1up&seq=6>
- Ayers RS, Westcot DW (1985) Water quality for agriculture. Irrigation and drainage paper 29. Food and Agriculture Organization of the United Nations: Rome. 174 p. <https://www.fao.org/3/t0234e/t0234e.pdf>
- Baer NW (1984) Nutrient content in ponderosa pine foliage: seasonal variation. Agricultural Experiment Station Technical Bulletins. 77. South Dakota State University. 10 p. http://openprairie.sdstate.edu/agexperimentsta_tb/77
- Bailey D, Bilderback T, Bir D (1999) Water considerations for container production of plants. NC State University Department of Horticultural Science Horticulture Information Leaflet 557: 11p. <https://www.clamerinforma.it/EnglishFiles/PDF/HIL557.pdf>
- Ballard R, Will GM (1978) Past and projected use of fertilisers in New Zealand forests. *NZJ For Sci* 8: 15-27. https://scion-web.squid.cloud/_data/assets/pdf_file/0019/59023/NZJFS811978BALLARD15_26.pdf
- Barbour HF, Berenyi NM (1969) Effect of liming on loblolly pine seedling growth and response to phosphorus application on organic soils in the greenhouse. Research Report 44. Westvaco, Forest Research. 16 p. <https://d.lib.ncsu.edu/collections/catalog/mc00496-002-bx0010-004-044#?c=&m=&s=&cv=&xywh=-2686%2C-373%2C10569%2C7448>
- Bates TE (1971) Factors affecting critical nutrient concentrations in plants and their evaluation: A review. *Soil Sci* 112(2): 116-130. <https://doi.org/10.1097/00010694-197108000-00005>
- Baule H (1975) World-wide use of fertilizer in forestry at present and in the near future. *South African Forestry Journal* 94(1): 13-19. <https://doi.org/10.1080/00382167.1975.9630443>
- Beaton JD, Moss A, MacRae I, Konkin JW, McGhee WPT, Kosick R (1965) Observations on foliage nutrient content of several coniferous tree species in British Columbia. *The Forestry Chronicle* 41(2) 222-236. <https://doi.org/10.5558/tfc41222-2>
- Bengtson GW (ed) (1968) Forest Fertilization-Theory and Practice. Tennessee Valley Authority, Muscle Shoals, AL: 316 p. <https://catalog.hathitrust.org/Record/001516717>
- Benzian B (1959) Nutrition problems in forest nurseries. *Science of Food and Agriculture* 10(12): 637-644. <https://doi.org/10.1002/jsfa.2740101201>
- Beyer WN, Green CE, Beyer M, Chaney RL (2013) Phytotoxicity of zinc and manganese to seedlings grown in soil contaminated by zinc smelting. *Environ Pollut* 179: 167-176.

- <https://doi.org/10.1016/j.envpol.2013.04.013>
- Bickelhaupt DH (1989) The long-term effect of a single application of horse manure on soil pH. *Tree Planters' Notes* 40(1): 31-33.
<https://rng.net/publications/tpn?SearchableText=Bickelhaupt+1989&formSubmitted=1>
- Binkley D, Fisher RF (2013) *Ecology and management of forest soils*. 4th ed. John Wiley & Sons.
<https://www.wiley.com/en-us/Ecology+and+Management+of+Forest+Soils%2C+4th+Edition-p-9781118422328>
- Binns WO, Mayhead GJ, MacKenzie JM (1980) Nutrient deficiencies of conifers in British forests. An illustrated guide. Leaflet 76. Forestry Commission. 23 p.
https://www.forestresearch.gov.uk/documents/7845/FCLF076_1980.pdf
- Blackmon BG (1969) Response of loblolly pine (*Pinus taeda* L.) seedlings to various levels and combinations of nitrogen and phosphorus. Baton Rouge, LA: Louisiana State University. 164 p. Ph.D. dissertation.
https://digitalcommons.lsu.edu/cgi/viewcontent.cgi?article=2637&context=gradschool_disstheses
- Blevins D, Allen HL, Colbert S, Gardner W (1996) Woodland owner notes: nutrition management for longleaf pinestraw. NCSU Coop. Ext. Service WON-30. Raleigh, NC: 8 p.
- Boardman R, Cromer RN, Lambert MJ, Webb MJ (1997) Forest plantations. In Reuter DJ, Robinson JB (eds.) *Plant analysis: an interpretation manual*. CSIRO Publishing, Melbourne:505-566.
- Bolton J, Benzian B (1970) Sulphur as a nutrient for Sitka spruce (*Picea sitchensis*) seedlings and radish (*Raphanus sativus*) grown on a sandy podzol in England. *The Journal of Agricultural Science* 74(3): 501-504. <https://doi.org/10.1017/S0021859600017615>
- Boxman AW, Krabbendam H, Bellemakers MJ, Roelofs JG (1991) Effects of ammonium and aluminium on the development and nutrition of *Pinus nigra* in hydroculture. *Environ Pollut* 73(2): 119-136.
[https://doi.org/10.1016/0269-7491\(91\)90018-R](https://doi.org/10.1016/0269-7491(91)90018-R)
- Boyer JN, South DB (1985) Nutrient content of nursery-grown loblolly pine seedlings. Circular 282. Alabama Agricultural Experiment Station, Auburn University, Auburn University, AL: 27 p.
<http://131.204.73.195/bitstream/handle/11200/2067/1279CIRC.pdf>
- Briggs RD (2008) Soils and nutrition: A forest nursery perspective. In: Dumroese RK; Riley LE, tech. coords. *National Proceedings: Forest and Conservation Nursery Associations-2007*. RMRS-P-57. USDA, Forest Service, Rocky Mountain Research Station, Fort Collins, CO: 55-64.
https://rng.net/publications/proceedings/2007/soils-and-nutrition-a-forest-nursery-perspective/at_download/file
- Bryson GM, Mills HA (eds) (2014) *Plant analysis handbook IV*. Micro-Macro Publishing: Athens, Georgia. 600 p. https://www.researchgate.net/publication/271849765_Plant_Analysis_Handbook_IV
- Bryson HL (1980) *Pisolithus tinctorius* mycobiont inoculations as a factor in performance of containerized and bare-root shortleaf pine seedlings on lignite minesoils in Panola County, Texas. DF thesis, Stephen F Austin State University, Nacogdoches. 418 p.
<https://scholarworks.sfasu.edu/cgi/viewcontent.cgi?article=1011&context=etds>
- Bücking H, Kuhn AJ, Schroeder WH, Heyser W (2002) The fungal sheath of ectomycorrhizal pine roots: an apoplastic barrier for the entry of calcium, magnesium, and potassium into the root cortex? *J Exp Bot* 53(374): 1659-1669. <https://doi.org/10.1093/jxb/erf011>
- Bueno SW, White EH, Bickelhaupt D (2012) Soil chemical properties in forest tree nurseries: conifer seedlings production. Lap Lambert Academic Publishing.
https://www.researchgate.net/publication/230744885_Soil_Chemical_Properties_in_Forest_Tree_Nurseries_Conifer_Seedlings_Production
- Bunting WR (1980) Seedling quality: growth and development—soil relationships, seedling growth and development, density control relationships. In: *Proceedings of the North American Forest Tree Nursery Soils Workshop*. Syracuse, NY: 21-42.
<https://babel.hathitrust.org/cgi/pt?id=pst.000007190872&view=1up&seq=9&skin=2021>
- Carey WA, South DB, Albrecht-Schmitt TE (2002) Gypsum crystals on roots of nursery-grown pine seedlings. *Commun Soil Sci Plan* 33(7-8): 1131-1137. <https://doi.org/10.1081/CSS-120003877>

- Carlson LW (1979) Guidelines for rearing containerized conifer seedlings in the Prairie Provinces. Information Report NOR-X-214. Environment Canada. Northern Forest Research Centre. Edmonton, Alberta: 62 p. <https://cfs.nrcan.gc.ca/publications?id=11650>
- Carrodus BB (1966) Absorption of nitrogen by mycorrhizal roots of beech: I. Factors affecting the assimilation of nitrogen. *New Phytologist* 65(3): 358-371. <https://nph.onlinelibrary.wiley.com/doi/pdf/10.1111/j.1469-8137.1966.tb06372.x>
- Carroll D (1962) Rainwater as a chemical agent of geologic processes: a review. US Government Printing Office, Washington, DC: 18 p. <https://citeseerx.ist.psu.edu/viewdoc/download?doi=10.1.1.832.5362&rep=rep1&type=pdf>
- Carter MR (1987) Seedling growth and mineral nutrition of Scots pine under acidic to calcareous soil conditions. *Soil Sci* 144(3): 175-180. <https://doi.org/10.1097/00010694-198709000-00003>
- Carter DR, Allen HL, Fox TR, Albaugh TJ, Rubilar RA, Campoe OC, Cook RL (2021) A 50-year retrospective of the Forest Productivity Cooperative in the southeastern United States: regionwide trials. *J Forest* 119(1): 73-85. <https://doi.org/10.1093/jofore/fvaa046>
- Chaganti VN, Culman SW, Herms C, Sprunger CD, Brock C, Soto AL, Doohan D (2021) Base cation saturation ratios, soil health, and yield in organic field crops. *Agron J* 113(5): 4190-4200. <https://doi.org/10.1002/agj2.20785>
- Chapman AG (1941) Tolerance of shortleaf pine seedlings for some variations in soluble calcium and H-ion concentration. *Plant Physiol* 16(2): 313-326. <https://doi.org/10.1104/pp.16.2.313>
- Chaves RDQ, Corrêa GF (2005) Macronutrients in the soil - *Pinus caribaea* Morelet system with yellowing of the needles followed by senescence and death. *Revista Árvore* 29: 691-700. <https://www.scielo.br/j/rarv/a/MpPkhHZrTPqGnNtxCZfKz7F/abstract/?lang=en>
- Christersson L (1973) The effect of inorganic nutrients on water economy and hardness of conifers. 1. The effect of varying potassium, calcium, and magnesium levels on water content, transpiration rate, and the initial phase of development of frost hardiness of *Pinus sylvestris* L. seedlings. *Studia Forestalia Suecica* 103: 1-26. <https://pub.epsilon.slu.se/5717/1/SFS103.pdf>
- Christersson L (1975) Frost-hardiness development in *Pinus sylvestris* L. seedlings at different levels of potassium and calcium fertilization. *Can J Forest Res* 5(4): 738-740. <https://doi.org/10.1139/x75-101>
- Coultas CL, Hsieh YP, McKee WH (1991) Loblolly pine seedling response to fertilizer and lime treatments on a spodosol. *Soil Sci Soc Am J* 55(3): 830-833. <https://doi.org/10.2136/sssaj1991.03615995005500030033x>
- Crannell WK, Tanaka Y, Myrold DD (1994) Calcium and pH interaction on root nodulation of nursery-grown red alder (*Alnus rubra* bong.) seedlings by Frankia. *Soil Biol Biochem* 26(5): 607-614. [https://doi.org/10.1016/0038-0717\(94\)90249-6](https://doi.org/10.1016/0038-0717(94)90249-6)
- Crowther EM (1950) Nutritional problems in forest nurseries. Report for 1949, Rothamsted Research: 122-129. <https://doi.org/10.23637/ERADOC-1-71>
- Cumming JR, Weinstein LH (1990) Aluminum-mycorrhizal interactions in the physiology of pitch pine seedlings. *Plant Soil* 125(1): 7-18. <https://doi.org/10.1007/BF00010739>
- Danielson RM (1966) The effect of soil fumigation on seedling growth, mycorrhizae and the associated microflora of loblolly pine (*Pinus taeda* L.) roots. MS thesis, North Carolina State University, Raleigh. 148 p.
- Davey CB (1988) Nursery soil management. In: Proceedings, southern forest nursery association: 80-85. https://rngr.net/publications/proceedings/1988/nursery-soil-management/at_download/file
- Davey CB (2002) Using soil test results to determine fertilizer applications. In: Dumroese RK, Riley LE, Landis TD (eds.) Proceedings, forest and conservation nursery associations-1999, 2000, and 2001. RMRS-P-24. USDA Forest Service, Rocky Mountain Research Station, Ogden UT: 22-26. <https://rngr.net/publications/proceedings/1999/davey.pdf>
- Davis AS, Jacobs DF, Wightman KE (2007a) Organic matter amendment of fallow forest tree seedling nursery soils influences soil properties and biomass of a sorghum cover crop. *Tree Planter's Notes* 52(1): 4-8. https://rngr.net/publications/tpn/52-1/organic-matter-amendment-of-fallow-forest-tree-seedling-nursery-soils-influences-soil-properties-and-biomass-of-a-sorghum-cover-crop/at_download/file

- Davis AS, Ross-Davis AL, Dumroese RK (2011) Nursery culture impacts cold hardiness in longleaf pine (*Pinus palustris*) seedlings. *Restor Ecol* 19(6): 717-719. <https://doi.org/10.1111/j.1526-100X.2011.00814.x>
- Davis M, Zue J, Clinton P (2015) Planted-forest nutrition. NZ For Res Inst Repot Info, Sheet 126 p. https://www.researchgate.net/profile/Jianming-Xue/publication/286931750_Planted_forest_Nutrition/links/5670f60508ae2b1f87acfb65/Planted-forest-Nutrition.pdf
- Davis MR, Coker G, Parfitt RL, Simcock R, Clinton PW, Garrett LG, Watt MS (2007b) Relationships between soil and foliar nutrients in young densely planted mini-plots of *Pinus radiata* and *Cupressus lusitanica*. *For Ecol Manag* 240(1-3): 122-130. https://www.academia.edu/download/45502190/Relationships_between_soil_and_foliar_nu20160510-32745-1h6icqg.pdf
- Deines J (1973) The effects of fertilization on the growth and development of 1-0 sycamore (*Platanus occidentalis* L.), sweetgum (*Liquidambar styraciflua* L.) and green ash (*Fraxinus pennsylvanica* Marsh.) seedlings. PhD thesis, North Carolina State University, Raleigh. 80 p
- Diaz DAR (2019) Chloride in Kansas: plant, soil, and fertilizer considerations. Kansas St. Univ. Ext. Bull. MF-2570. Kansas State University, KS: 1-4. <https://bookstore.ksre.ksu.edu/pubs/MF2570.pdf>
- Dickens ED, Clabo DC, Moorhead D (2021) Longleaf pine wood yields response to midrotation fertilization on two old-field sites. WFSNR-21-72A. University of Georgia, Athens, GA: 13 p. <https://bugwoodcloud.org/resource/files/21158.pdf>
- Dickson A, Leaf AL, Hosner JF (1960) Seedling quality - soil fertility relationships of white spruce, and red and white pine in nurseries. *The Forestry Chronicle* 36(3): 237-241. <https://pubs.cif-ifc.org/doi/pdf/10.5558/tfc36237-3>
- Dobrahner J, Lowery B, Iyer JG (2004) Slow-release fertilization reduces nitrate leaching in bareroot production of *Pinus strobus* seedlings. In: Riley LE, Dumroese RK, Landis TD (eds.) National proceedings: Forest and Conservation Nursery Associations. RMRS-P-33. USDA Forest Service, Rocky Mountain Research Station. Fort Collins, CO: 129-139. https://rngr.net/publications/proceedings/2003/PDF.2004-06-08.4503/at_download/file
- Donald DGM (1991) Nursery fertilization of conifer planting stock. In: van den Driessche R (ed) *Mineral Nutrition of Conifer Seedlings*: 135-167.
- Donald DGM, Young I (1982) The growth of pine seedlings in South African forest nurseries. *South African Forestry Journal* 123(1): 36-50. <https://doi.org/10.1080/00382167.1982.9628852>
- Dumroese RK, Wenny DL (1997) Fertilizer regimes for container-grown conifers of the Intermountain West. In Haase DL, Rose R (ed) *Forest seedling nutrition from the nursery to the field*: 28-29. https://rngr.net/publications/proceedings/1997/fertilizer-regimes-for-container-grown-conifers-of-the-intermountain-west/at_download/file
- Eberhardt PJ, Pritchett WL (1971) Foliar applications of nitrogen to slash pine seedlings. *Plant Soil* 34(1): 731-739. <https://doi.org/10.1007/BF01372827>
- Erdmann GG (1966) Promising conifers for western Iowa. Research Paper NC-8. USDA Forest Service. North Central Forest Experiment Station. Marquette (MN): 8 p.
- Flaten CM (1939) A study on chlorosis of *Pinus resinosa* in a forest nursery, with special reference to some mineral deficiencies. MF thesis. University of Michigan, Ann Arbor: 35 p. <https://deepblue.lib.umich.edu/bitstream/handle/2027.42/114430/39015003261644.pdf?sequence=2>
- Flinn DW, Homans P, Craig FG (1980) Survey of the nutrient status of *Pinus radiata* seedlings and of soil properties in three Victorian nurseries. *Aust Forestry* 43(1): 58-66. <https://doi.org/10.1080/00049158.1980.10674246>
- Flinn DW, Waugh RJ (1983) Evaluation of gypsum and organic matter additions for improving soil structure in a radiata pine nursery at Benalla, Victoria. *Australian Journal of Experimental Agriculture* 23(121): 208-215. <https://doi.org/10.1071/EA9830208>
- Fowells HA, Krauss RW (1959) The inorganic nutrition of loblolly pine and Virginia pine with special reference to nitrogen and phosphorus. *Forest Science* 5(1): 95-112. <https://doi.org/10.1093/forestscience/5.1.95>

- Fox WF (1904) Forest nurseries and nursery methods in Europe. JB Lyon Company, Albany. <https://babel.hathitrust.org/cgi/pt?id=loc.ark:/13960/t68348s41&view=1up&seq=3&skin=2021>
- Franklin JA, Zwiazek JJ, Renault S, Croser C (2002) Growth and elemental composition of jack pine (*Pinus banksiana*) seedlings treated with sodium chloride and sodium sulfate. *Trees* 16(4): 325-330. <https://www.academia.edu/download/54065916/s00468-002-0175-520170805-15730-18v93l9.pdf>
- Fried M, Peech M (1946) Comparative effects of lime and gypsum upon plants grown on acid soils. *Journal of the American Society of Agronomy*. 38(7): 614-623. <https://doi.org/10.2134/agronj1946.00021962003800070004x>
- Gaspar AP, Laboski CAM (2016) Base saturation: What is it? Should I be concerned? Does it affect my fertility program? In: *Proceedings Wis. Crop Manage. Conf* 5: 55-61. <https://extension.soils.wisc.edu/wp-content/uploads/sites/68/2014/02/WCMC-2016-Complete-Proceedings.pdf#page=63>
- Gleason JF (1989) Fertilization of 2-0 ponderosa pine seedlings in the nursery and field: morphology, physiology, and field performance. MS thesis, Oregon State University, Corvallis. 109 p. <https://ir.library.oregonstate.edu/downloads/5712mb473>
- Goodrich BA, Jacobi WR (2012) Foliar damage, ion content, and mortality rate of five common roadside tree species treated with soil applications of magnesium chloride. *Water, Air, & Soil Pollution* 223(2): 847-862. <https://doi.org/10.1007/s11270-011-0907-5>
- Goslin WE (1959) Effects of deficiencies of essential elements on the development and mineral composition of seedlings of Scots pine (*Pinus sylvestris* L.). PhD thesis, The Ohio State University. Columbus. 114 p. https://etd.ohiolink.edu/apexprod/rws_etd/send_file/send?accession=osu1486474943922309&disposition=inline
- Göttlein A (2015) Grenzwertbereiche für die ernährungsdiagnostische Einwertung der Hauptbaumarten Fichte, Kiefer, Eiche, Buche. *AllgForstJagdztg* 186(5/6): 110–116. https://www.waern.wzw.tum.de/fileadmin/media/waldernaehrung/Goettlein_Grenzwertbereiche_fuer_die_ernaehrungsdiagnostische_Einwertung_der_Haupbaumarten_AFJZ_2015.pdf
- Gregoire N, Fisher RF (2004) Nutritional diagnoses in loblolly pine (*Pinus taeda* L.) established stands using three different approaches. *For Ecol Manag* 203(1-3): 195-208. <https://doi.org/10.1016/j.foreco.2004.07.049>
- Gupta UC, MacLeod JA (1981) Plant and soil boron as influenced by soil pH and calcium sources on podzol soils. *Soil Sci* 131(1): 20-25. <https://doi.org/10.1097/00010694-198101000-00003>
- Hachani C, Lamhamedi MS, Cameselle C, Gouveia S, Zine El Abidine A, Khasa DP, Béjaoui Z (2020) Effects of ectomycorrhizal fungi and heavy metals (Pb, Zn, and Cd) on growth and mineral nutrition of *Pinus halepensis* seedlings in North Africa. *Microorganisms* 8(12): 2033. <https://www.mdpi.com/2076-2607/8/12/2033/htm>
- Hacskaylo J, Finn RF, Vimmerstedt JP (1969) Deficiency symptoms of some forest trees. *Research Bulletin* 1015. Ohio Agricultural Research and Development Center, Wooster, OH: 69 p. https://kb.osu.edu/bitstream/handle/1811/62823/1/OARDC_research_bulletin_n1015.pdf
- Hallett RA, Hornbeck JW (1997) Foliar and soil nutrient relationships in red oak and white pine forests. *Can J Forest Res* 27(8): 1233-1244. <https://doi.org/10.1139/x97-026>
- Hallet RD (1980) Experience in the use of soil analysis data. In: *Proceedings of the North American Forest Tree Nursery Soils Workshop*. Syracuse, NY: 296-298. <https://babel.hathitrust.org/cgi/pt?id=pst.000007190872&view=1up&seq=9&skin=2021>
- Hans RR (2013) Initial growth responses to controlled release fertilizer application at establishment of commercial forestry species in South Africa. PhD thesis, Stellenbosch University. Stellenbosch. 158 p. <http://scholar.sun.ac.za/handle/10019.1/85609>
- Hansen TS (1923) Use of fertilizers in a coniferous nursery. *J Forest* 21(7): 732-735. <https://doi.org/10.1093/jof/21.7.732>
- Hart PBS, Widdowson JP (1981) The response of caribbean pine, green panic, and siratro to fertiliser on soils of the 'Eua Uplands, Tonga. *New Zealand Journal of Experimental Agriculture* 9(3-4): 255-262. <https://doi.org/10.1080/03015521.1981.10425423>

- Hawkesford M, Horst W, Kichey T, Lambers H, Schjoerring J, Møller IS, White P (2012) Functions of macronutrients. In: Marschner's mineral nutrition of higher plants. Academic Press: 135-189.
- Heckman JR, Sims JT, Beegle DB, Coale FJ, Herbert SJ, Bruulsema TW, Bamka WJ (2003) Nutrient removal by corn grain harvest. *Agron J* 95(3): 587-591. <https://doi.org/10.2134/agronj2003.5870>
- Heidmann LJ, Thorud DB (1976) Controlling frost heaving of ponderosa pine seedlings in Arizona. RM-172. USDA Forest Service, Rocky Mountain Forest and Range Experiment Station, Fort Collins, CO: 12 p. <https://www.biodiversitylibrary.org/item/177590#page/3/mode/1up>
- Helm CW, Kuser JE (1991) Container growing pitch pine: germination, soil pH, and outplanting size. *North J Appl For* 8(2): 63-68. <https://doi.org/10.1093/njaf/8.2.63>
- Helmisaari HS (1992) Nutrient retranslocation within the foliage of *Pinus sylvestris*. *Tree Physiol* 10(1): 45-58. <https://doi.org/10.1093/treephys/10.1.45>
- Hinesley LE, Maki TE (1980) Fall fertilization helps longleaf pine nursery stock. *South J Appl Forest* 4(3): 132-135. <https://doi.org/10.1093/sjaf/4.3.132>
- Holopainen JK, Rikala R, Kainulainen P, Oksanen J (1995) Resource partitioning to growth, storage and defence in nitrogen-fertilized Scots pine and susceptibility of the seedlings to the tarnished plant bug *Lygus rugulipennis*. *New Phytologist* 131(4): 521-532. <https://doi.org/10.1111/j.1469-8137.1995.tb03088.x>
- Hoyle MC, Mader DL (1964) Relationships of foliar nutrients to growth of red pine in western Massachusetts. *Forest Sci* 10(3): 337-347. <https://academic.oup.com/forestscience/article-abstract/10/3/337/4746265?redirectedFrom=fulltext>
- Humphreys FR (1964) The nutrient status of pine plantations in central New South Wales. *Appita* 18(11): 111-120.
- Hunter IR (1996) The occurrence and treatment of magnesium deficiency in radiata pine in New Zealand. New Zealand Forest Research Institute, FRI Bulletin No. 172. 136 p. <https://scion.contentdm.oclc.org/digital/api/collection/p20044coll6/id/166/download>
- Ingestad T (1962) Macro element nutrition of pine, spruce, and birch seedlings in nutrient solutions. *Meddelandenfrån Statensskogsforsknings institut* 51(7): 1-154. https://pub.epsilon.slu.se/10053/1/medd_statens_skogsforskningsinst_051_07.pdf
- Ingestad T (1974) Towards optimum fertilization. *Ambio* 3(2): 49-54. <https://www.jstor.org/stable/4312045>
- Iyer JG, Schulte EE, Randall GW (1971) Relationship between foliar composition of red-pine and jack-pine seedlings and vulnerability to *Lophodermium* needle-cast disease. *Plant Soil* 35(1): 213-215. <https://link.springer.com/content/pdf/10.1007/BF01372651.pdf>
- Januszek K, Stępniewska H, Błońska E, Molicka J, Koziel K, Gdula A, Wójs A (2014) Impact of aluminum sulphate fertilizer on selected soil properties and the efficiency and quality of pine seedlings in the forest ground tree nursery. *Leśne Prace Badawcze* 75(2): 127-138. <https://depot.ceon.pl/bitstream/handle/123456789/5193/doi-10-2478-frp-2014-0012%20e.pdf?sequence=1&isAllowed=y>
- Jenkinson JL (1974) Ponderosa pine progenies: differential response to ultramafic and granitic soils. PSW-101. USDA, Forest Service, Pacific Southwest Forest and Range Experiment Station. Berkeley, CA: 14 p. <https://www.fs.usda.gov/treesearch/pubs/28658>
- Jokela EJ (2004) Nutrient management of southern pines. In: Dickens ED, Barnett JP, Hubbard WG, Jokela EJ (eds) *Slash pine: still growing and growing*. GTR-SRS-76. USDA Forest Service, Southern Research Station, Asheville, NC: 27-35. https://www.srs.fs.usda.gov/pubs/gtr/gtr_srs076.pdf
- Jones GW (1925) Forest nursery working practice at Savenac Nursery. *J Forest* 23(7): 635-644. <https://doi.org/10.1093/jof/23.7.635>
- Jose S, Merritt S, Ramsey CL (2003) Growth, nutrition, photosynthesis and transpiration responses of longleaf pine seedlings to light, water and nitrogen. *Forest Ecol Manag* 180(1-3): 335-344. [https://doi.org/10.1016/S0378-1127\(02\)00583-2](https://doi.org/10.1016/S0378-1127(02)00583-2)
- Kavvadias VA (1996) Manganese and calcium interaction on the growth and nutrient uptake of Scots pine and black pine seedlings. Aberdeen, SCT: University of Aberdeen. 635 p. Ph.D. dissertation. <https://www.proquest.com/docview/301456399?pq-origsite=gscholar&fromopenview=true>
- Kavvadias VA, Miller HG (1999) Manganese and calcium nutrition of *Pinus sylvestris* and *Pinus nigra* from two different origins. II. Calcium. *Forestry* 72(2): 147-156.

- <https://doi.org/10.1093/forestry/72.2.147>
- Kieliszewska-Rokicka B (1991) Effect of ammonium and nitrate nutrition on hydrolytic enzymes activity of Scots pine (*Pinus sylvestris* L.) roots and phosphorus content in shoots. *Arboretum Kórnickie* 36: 127-136. https://rcin.org.pl/id/Content/159472/PDF/KOR001_149004.pdf
- Kishchuk BE (2000) Calcareous soils, their properties and potential limitations to conifer growth in Southeastern British Columbia and Western Alberta: a literature review. Information Report NOR-X-370, Northern Forestry Centre, Canadian Forest Service, Edmonton, AB: 21 p. <https://publications.gc.ca/site/eng/9.560988/publication.html>
- Knight PJ (1978a) Fertilizer practice in New Zealand forest nurseries. *NZJ For Sci* 8(1): 27-53. https://www.scionresearch.com/_data/assets/pdf_file/0018/59022/NZJFS811978KNIGHT27_53.pdf
- Knight PJ (1978b) The nutrient content of *Pinus radiata* seedlings: a survey of planting stock from 17 New Zealand forest nurseries. *NZJ For Sci* 8(1): 54-69. https://www.scionresearch.com/_data/assets/pdf_file/0006/37185/NZJFS811978KNIGHT54_69.pdf
- Knight PJ (1981) The maintenance of productivity in forest nurseries. In: FRI Symposium 22. New Zealand Forest Service, Forest Research Institute: 48-69.
- Kopittke PM, Menzies NW (2007) A review of the use of the basic cation saturation ratio and the "ideal" soil. *Soil Sci Soc Am J* 71(2): 259-265. <http://www.agvise.com/wp-content/uploads/2015/02/SSSA-Cation-Ratios.pdf>
- Kormanik PP, Sung SJS, Kormanik TL (1994) Irrigating and fertilizing to grow better nursery seedlings. In: Landis TD (ed). Proceedings: northeastern and intermountain forest and conservation nursery associations. GTR-RM-243. USDA Forest Service, Rocky Mountain Forest and Range Experiment Station, Fort Collins, CO: 115-121. https://rngr.net/publications/proceedings/1993/kormanik.pdf/at_download/file
- Kumar D, Shivay YS (2008) Definitional Glossary of Agricultural Terms: Volume I. IK International Publishing House. New Delhi. 314 p.
- Kyle KH, Andrews LJ, Fox TR, Aust WM, Burger JA (2005) Long-term effects of drainage, bedding, and fertilization on growth of loblolly pine (*Pinus taeda* L.) in the Coastal Plain of Virginia. *South J Appl Forest* 29(4): 205-214. <https://doi.org/10.1093/sjaf/29.4.205>
- Landis TD (1979) The saline soil syndrome and its effect on bare-root production in two Rocky Mountain Area nurseries. In: Proceedings of Intermountain Nurseryman's Association Meeting. USDA Forest Service, Carbondale, CO: 78-81. https://rngr.net/publications/proceedings/1979/PDF.2004-01-30.3713/at_download/file
- Landis TD (1988) Management of forest nursery soils dominated by calcium salts. *New Forest* 2(3): 173-193. <https://doi.org/10.1007/BF00029987>
- Landis TD (1996) Secondary nutrients - calcium. *Forest Nursery Notes* 16(1): 8-11. https://rngr.net/publications/fnn/1996-winter/articles/secondary-nutrients2014calcium/at_download/file
- Landis TD, Tinus RW, McDonald SE, Barnett JP (1989) Seedling nutrition and irrigation. In: The container tree nursery manual. Agricultural Handbook 674, Volume 4. USDA Forest Service, Washington, DC: 119 p. <https://rngr.net/publications/ctnm/volume-4>
- Landis TD, Pinto JR, Davis AS (2009) Fertigation-injecting soluble fertilizers into the irrigation system. *Forest Nursery Notes* 29(2): 4-13. <https://www.fs.usda.gov/rmrs/publications/fertigation-injecting-soluble-fertilizers-irrigation-system>
- Larsen HS, South DB, Boyer JN (1988) Foliar nitrogen content at lifting correlates with early growth of loblolly pine seedlings from 20 nurseries. *South J Appl Forest* 12(3): 181-185. <https://doi.org/10.1093/sjaf/12.3.181>
- Leski T, Aučina A, Skridaila A, Pietras M, Riepšas E, Rudawska M (2010) Ectomycorrhizal community structure of different genotypes of Scots pine under forest nursery conditions. *Mycorrhiza* 20(7): 473-481. <https://doi.org/10.1007/s00572-010-0298-2>
- Lipman CB (1916) A critique of the hypothesis of the lime-magnesia ratio. *The Plant World* 19(4): 83-105.

- <https://www.jstor.org/stable/pdf/43477507.pdf>
- Lunt HA (1938) The use of fertilizers in the coniferous nursery. Bulletin 416, Connecticut Agricultural Experiment Station: New Haven CT: 721-766. <https://portal.ct.gov/-/media/CAES/DOCUMENTS/Publications/Bulletins/B416pdf.pdf>
- Lunt HA (1947) The response of hybrid poplar and other forest tree species to fertilizer and lime treatment in concrete soil frames. Journal of Agricultural Research 74(4): 113-132.
- Lyle ES (1969) Mineral deficiency symptoms in loblolly pine seedlings. Agron J 61(3): 395-398. <https://doi.org/10.2134/agronj1969.00021962006100030019x>
- Lyle ES, Adams F (1971) Effect of available soil calcium on taproot elongation of loblolly pine (*Pinus taeda* L.) seedlings. Soil Sci Soc Am J 35(5): 800-805. <https://doi.org/10.2136/sssaj1971.03615995003500050046x>
- Lyle ES, Pearce ND (1968) Sulfur deficiency in nursery seedlings may be caused by concentrated fertilizers. Tree Planters' Notes 19(1): 9-10. https://rngr.net/publications/tpn/19-1/pdf.2005-05-20.5572607495/at_download/file
- Madgwick HAI, Ovington JD (1959) The chemical composition of precipitation in adjacent forest and open plots. Forestry 32(1): 14-22. <https://doi.org/10.1093/forestry/32.1.14>
- Madgwick HAI (1964) The chemical composition of foliage as an index of nutritional status in red pine (*Pinus resinosa* Ait). Plant Soil 21(1): 70-80. <https://doi.org/10.1007/BF01373874>
- Maki TE, Henry BW (1951) Root-rot control and soil improvement at the Ashe Forest Nursery. Occasional Paper 119. USDA Forest Service, Southern Forest Experiment Station. New Orleans, LA: 23 p. <https://archive.org/details/CAT31363714/page/16/mode/2up>
- Malavolta E, Sarruge JR, Haag HP, Vencovsky R, Santos CFO, Valsechi O, Scoton LC, Coelho RSG (1970) The relation of the concentration of macronutrients in the substrate and in the foliage to cell wall thickness and cellulose concentration in the xylem of slash pine (*Pinus elliottii*). Anais da Escola Superior de Agricultura Luiz de Queiroz 27: 295-333. <https://www.scielo.br/pdf/aesalq/v27/22.pdf>
- Marschner H (2012) Marschner's mineral nutrition of higher plants. Academic press. 684 p. <https://doi.org/10.1016/C2009-0-63043-9>
- Martian BF (1989) Soil management practices at the Big Sioux Nursery. In: Landis TD (ed) Proceedings, Intermountain Forest Nursery Association. GTR RM-184. USDA Forest Service, Rocky Mountain Forest and Range Experiment Station: 82-85. https://rngr.net/publications/proceedings/1989/martian.pdf/at_download/file
- Marx DH (1990) Soil pH and nitrogen influence *Pisolithus* ectomycorrhizal development and growth of loblolly pine seedlings. Forest Sci 36(2): 224-245. <https://doi.org/10.1093/forestscience/36.2.224>
- Marx DH, Cordell CE, Kenney DS, Mexal JG, Artman JD, Riffle JW, Molina RJ (1984) Commercial vegetative inoculum of *Pisolithus tinctorius* and inoculation techniques for development of ectomycorrhizae on bare-root tree seedlings. For Sci 30(3): Monograph 25. https://academic.oup.com/forestscience/article-abstract/30/suppl_1/a0001/4656736
- May B, Smethurst P, Carlyle C, Mendham D, Bruce J, Baillie C (2009) Review of fertiliser use in Australian forestry. Forest and Wood Products Australia Report PRC072-0708. 96 p. https://www.fwpa.com.au/images/processing/PRC072-0708_Fertiliser_Review_Research_Report_0.pdf
- May JT, Johnson HH, Gilmore AR (1962) Chemical composition of southern pine seedlings. Research Paper 10. Georgia Forest Research Council, Macon, GA: 11 p.
- May JT (1984) Nutrients and fertilization. In: Lantz CW ed: Southern Pine Nursery Handbook. Atlanta, GA: USDA Forest Service, Southern Region: 1201-1241. https://rngr.net/publications/spnh/PDF.2003-08-12.3705/at_download/file
- Maxwell GR (2012) Synthetic nitrogen products. In: Handbook of Industrial Chemistry and Biotechnology: 875-937. https://doi.org/10.1007/978-1-4614-4259-2_22
- McConnell RC, Klages MG (1969) Forest nursery soils of northern Idaho and western Montana. Montana Agricultural Experiment Station Montana State University, Bozeman. 33 p. <https://babel.hathitrust.org/cgi/pt?id=uiug.30112019700563&view=1up&seq=6&skin=2021>

- McIntyre AC, White JW (1930) The growth of certain conifers as influenced by different fertilizer treatments. *Agron J* 22(6): 558-567. <https://doi.org/10.2134/agronj1930.00021962002200060012x>
- McKee WH (1978) Slash pine seedling response to potassium and calcium on imperfectly drained coastal plain soil. *Plant Soil* 50(1): 615-624. <https://doi.org/10.1007/BF02107213>
- McLeod KW, Sherrod C, Porch TE (1979) Response of longleaf pine plantations to litter removal. *For Ecol Manag* 2: 1-12. [https://doi.org/10.1016/0378-1127\(79\)90032-x](https://doi.org/10.1016/0378-1127(79)90032-x)
- McNabb K, Heidbreder-Olson E (1998) Results of the 1996 irrigation water quality survey. Research report 98-05. Auburn, AL: Auburn University, Southern Forest Nursery Management Cooperative. 7 p.
- Mellert KH, Göttelein A (2012) Comparison of new foliar nutrient thresholds derived from van den Burg's literature compilation with established central European references. *Eur J Forest Res* 131(5): 1461-1472. <https://link.springer.com/content/pdf/10.1007/s10342-012-0615-8.pdf>
- Menzies MI, Holden DG, Green LM, Rook DA (1981) Seasonal changes in frost tolerance of *Pinus radiata* seedlings raised in different nurseries. *NZJ For Sci* 11(2): 100-111. https://scion-web.squidcloud.com/data/assets/pdf_file/0009/59319/NZJFS1121981MENZIES100_111.pdf
- Metz LJ, Wells CG, Swindel BF (1966) Sampling soil and foliage in a pine plantation. *Soil Science Society of America Journal* 30(3): 397-399. <https://doi.org/10.2136/sssaj1966.03615995003000030027x>
- Mexal JG, Fisher JT (1987) Organic matter amendments to a calcareous forest nursery soil. *New Forest* 1(4): 311-323. <https://doi.org/10.1007/BF00031741>
- Mitchell HL (1939) The growth and nutrition of white pine (*Pinus strobus* L.) seedlings in cultures with varying nitrogen, phosphorus, potassium and calcium: with observations on the relation of seed weight to seedling yield. *Black Rock Forest Bulletin* 9, Cornwall on the Hudson, NY: 135 p. https://www.blackrockforest.org/wp-content/uploads/2021/03/brf_bulletin_9.pdf
- Mitchell RJ, Garrett HE, Cox GS, Atalay A (1990) Boron and ectomycorrhizal influences on mineral nutrition of container-grown *Pinus ehinata* mill. *J Plant Nutr* 13(12): 1555-1574. <https://doi.org/10.1080/01904169009364175>
- Morris WG (1979) Comparison of sulfur coated urea, nitroform and ammonium sulfate as a source of nitrogen for loblolly pine nursery production. Weyerhaeuser Co. Tech. Rep. 042-2008/79/1.
- Moschler WW, Jones GD, Adams RE (1970) Effects of loblolly pine fertilization on a piedmont soil: growth, foliar composition, and soil nutrients 10 years after establishment. *Soil Sci Soc Am J* 34(4): 683. <https://doi.org/10.2136/sssaj1970.03615995003400040039x>
- Moser F (1933) The calcium-magnesium ratio in soils and its relation to plant growth. *Jour Am Soc Agron* 25: 365-377. <https://doi.org/10.2134/agronj1933.00021962002500060001x>
- Munson KR (1982) Decomposition, function, and maintenance of organic matter in a sandy nursery soil. PhD thesis, University of Florida, Gainesville. 96 p. <http://file.iflora.cn/fastdfs/group1/M00/64/46/wKhnoF2NwXSAMM8tAD0yVvJ61sI354.pdf>
- Murison WF (1960) Macronutrient deficiency and its effect on coniferous growth. PhD thesis, University of British Columbia, Vancouver. 235 p. <https://open.library.ubc.ca/collections/831/831/items/1.0106016>
- Mylavarapu RS, Sanchez JF, Nguyen JH, Bartos JM (2002) Evaluation of Mehlich-1 and Mehlich-3 extraction procedures for plant nutrients in acid mineral soils of Florida. *Commun Soil Sci Plan* 33(5-6): 807-820. <https://doi.org/10.1081/CSS-120003067>
- Nakos G (1979) Lime-induced chlorosis in *Pinus radiata*. *Plant Soil* 52(4): 527-536. <https://doi.org/10.1007/BF02277948>
- Nelson LE, Switzer GL (1985) Trends in the maintenance of soil fertility in Mississippi Nurseries. In: South DB (ed.). *Proc. of the International Symposium on Nursery Management for the Southern Pines*. Auburn University, AL: 222-236.
- Nicol W (1820) *The planter's kalendar; or the nurseryman's & forester's guide in the operations of the nursery, the forest, and the grove*. Archibald Constable and Company. <https://catalog.hathitrust.org/Record/O11537915>
- North Carolina State Forest Nutrition Cooperative (NCSFNC) (1991) Descriptive statistics and relationships among soil and foliar characteristics in midrotation loblolly pine plantations. Res. Note 7. College of Forest Resources, North Carolina State University, Raleigh, NC: 29 p.

- Olykan ST, Adams JA, Nordmeyer AH, McLaren RG (1995) Micronutrient and macronutrient uptake by *Pinus radiata*, and soil boron fractions, as affected by added nitrogen and boron. *NZ J Forestry Sci* 25(1): 61-72. https://www.scionresearch.com/_data/assets/pdf_file/0008/59633/NZJFS2511995OLYKAN61-72.pdf
- O'Reilly C, De Atrip N, Doody C, O'Reilly D, Doody P, Thompson B (2008) Increasing the yield and quality of broadleaf planting stock through higher N fertilisation in the nursery. *Irish Forestry* 65(1-2): 5-16. <https://journal.societyofirishforesters.ie/index.php/forestry/article/view/10006>
- Pellett HM, Carter JV (1981) Effect of nutritional factors on cold hardiness of plants. *Horticultural Reviews* 3: 144-171. <https://doi.org/10.1002/9781118060766.ch4>
- Percival G, Barnes S (2008) Calcium-induced freezing and salinity tolerance in evergreen oak and apple cv. 'Golden Crown'. *Arboriculture and Urban Forestry* 34(3): 191-199. <https://doi.org/10.48044/jauf.2008.026>
- Pessin LJ (1937) The effect of nutrient deficiency on the growth of longleaf pine seedlings. USDA Forest Service, Southern Forest Experiment Station. New Orleans LA: Occasional Paper 65: 1-7. <https://babel.hathitrust.org/cgi/pt?id=mdp.39015003619577&view=1up&seq=325&skin=2021&q1=peessin>
- Pharis RP, Barnes RL, Naylor AW (1964) Effects of nitrogen level, calcium level and nitrogen source upon the growth and composition of *Pinus taeda* L. *Physiol Plantarum* 17(3): 560-572. <https://doi.org/10.1111/j.1399-3054.1964.tb08185.x>
- Potvin LR, Jurgensen MF, Dumroese RK, Richter DL, Page-Dumroese DS (2014) Mosaic stunting in bareroot *Pinus banksiana* seedlings is unrelated to colonization by mycorrhizal fungi. *New Forest* 45(6): 893-903.
- Powers RF (1983) Forest fertilization research in California. In: Ballard R, Gessel SP (eds). IUFRO symposium on forest site and continuous productivity. GTR-PNW-163. USDA Forest Service, Pacific Northwest Research Station, Portland, OR: 388-397. <https://www.fs.usda.gov/treearch/pubs/7571>
- Prado RM (2021) Mineral nutrition of tropical plants. Springer, Cham. <https://doi.org/10.1007/978-3-030-71262-4>
- Pritchett WL, Comerford (1983) Nutrition and fertilization of slash pine. In: The Managed Slash Pine Ecosystem. Stone EL (ed.). University of Florida, Gainesville, FL: 69-90. <https://agris.fao.org/agris-search/search.do?recordID=US8475671>
- Purnell HM (1958) Nutritional studies of *Pinus radiata* Don: 1. Symptoms due to deficiencies of some major elements. *Aust Forestry* 22(2): 82-87. <https://doi.org/10.1080/00049158.1958.10675852> <https://doi.org/10.1080/1065657X.2004.10702158>
- Retan GA (1914) Effective fertilizers in nurseries. *J Forestry* 12(1): 34-36. <https://doi-org.spot.lib.auburn.edu/10.1093/jof/12.1.34>
- Richards BN (1965) Mycorrhiza development of loblolly pine seedlings in relation to soil reaction and the supply of nitrate. *Plant Soil* 22(2): 187-199. <https://doi.org/10.1007/BF01373990>
- Richards BN, Wilson GL (1963) Nutrient supply and mycorrhiza development in Caribbean pine. *Forest Sci* 9(4): 405-412. <https://doi.org/10.1093/forestscience/9.4.405>
- Richardson KF, Perkins RW (1985) Lesotho woodlot project, *Pinus radiata* nursery nutrition experiment. *The Commonwealth Forestry Review* 64(3): 267-280. <https://www.jstor.org/stable/42608052>
- Ritchey KD, Silva JE, Costa UF (1982) Calcium deficiency in clayey B horizons of savanna oxisols. *Soil Science* 133(6): 378-382. <https://doi.org/10.1097/00010694-198206000-00007>
- Rocha JHT, du Toit B, Goncalves JLM (2019) Ca and Mg nutrition and its application in Eucalyptus and *Pinus* plantations. *For Ecol Manag* 442: 63-78. <https://doi.org/10.1016/j.foreco.2019.03.062>
- Rosendahl R, Korstian CF (1945) Effect of fertilizers on loblolly pine in a North Carolina nursery. *Plant Physiology* 20(1): 19-23. <https://www.ncbi.nlm.nih.gov/pmc/articles/PMC437694/pdf/plntphys00275-0028.pdf>
- Rowan SJ (1971) Soil fertilization, fumigation, and temperature affect severity of black root rot of slash pine. *Phytopathology* 61: 184-187.

- https://www.apsnet.org/publications/phytopathology/backissues/Documents/1971Articles/Phyto61n02_184.pdf
- Sadreika V (1976) Processing soil and plant data print-outs and recommendations. In: Proceedings, Northeastern Area Nurserymen's Conference, Kemptville, Ontario: 27-46. https://rngr.net/publications/proceedings/1976/processing-soil-and-plant-data-print-outs-and-recommendations/at_download/file
- Sands R, Clarke ARP (1977) Response of radiata pine to salt stress. I. Water relations, osmotic adjustment and salt uptake. Aust J Plant Physiol 4(4): 637-646. <https://doi.org/10.1071/PP9770637>
- Saure MC (2014) Why calcium deficiency is not the cause of blossom-end rot in tomato and pepper fruit—a reappraisal. Scientia Horticulturae 174: 151-154. <https://doi.org/10.1016/j.scienta.2014.05.020>
- Schaberg PG, DeHayes DH, Hawley GJ, Strimbeck GR, Cumming JR, Murakami PF, Borer CH (2000) Acid mist and soil Ca and Al alter the mineral nutrition and physiology of red spruce. Tree Physiol 20(2): 73-85. <https://doi.org/10.1093/treephys/20.2.73>
- Schenck CA (1907) Biltmore lectures on silviculture. Brandow Printing Company, NY. <https://www.biodiversitylibrary.org/item/59977#page/7/mode/1up>
- Scovell MA (1890) Commercial fertilizers. Kentucky Agricultural Experiment Station, Bulletin 29. <https://babel.hathitrust.org/cgi/pt?id=uiug.30112056313296&view=1up&seq=1&skin=2021>
- Shorrocks VM (1997) The occurrence and correction of boron deficiency. Plant Soil 193(1): 121-148. <https://doi.org/10.1023/A:1004216126069>
- Schulte EE, Kelling KE (1985) Soil Calcium to Magnesium Ratios--should You be Concerned? A2986. University of Wisconsin, Madison: 4 p. <http://corn.agronomy.wisc.edu/Management/pdfs/a2986.pdf>
- Shoulders E, Czabator FJ (1965) Chlorosis in a southern pine nursery: a case study. Tree Planters' Notes 71(1): 19-21. <https://rngr.net/publications/tpn/16-2>
- Show SB (1930) Forest nursery and planting practice in the California pine region. Circular 92. USDA, Washington, DC: 75 p. <https://babel.hathitrust.org/cgi/pt?id=uiug.30112019276846&view=1up&seq=1&skin=2021>
- Simpson JA (1985) Use of inorganic fertilizers and cover crops in exotic pine nurseries of southern Queensland, Australia. In: South DB (ed.). Proc. of the International Symposium on Nursery Management for the Southern Pines. Auburn University, AL: 203-212. <https://nurserycoop.auburn.edu/PDF%20files/organic2.pdf>
- Slaton SH, Iyer JG (1974) Manganese compounds harmful to planting stock under some soil conditions. Tree Planters' Notes 25(2): 19-21. <https://rngr.net/publications/tpn/25-2/PDF.2003-09-25.2347/>
- Smits MM, Wallander H (2017) Role of mycorrhizal symbiosis in mineral weathering and nutrient mining from soil parent material. In: Mycorrhizal Mediation of Soil: 35-46. <https://doi.org/10.1016/B978-0-12-804312-7.00003-6>
- Snowdon P, Waring HD (1985) Effects of factorial combinations of urea, dicalcium phosphate, gypsum, and potassium chloride on growth and foliage composition of closely spaced *Pinus radiata*. Australian Forest Research 15: 333-352.
- Solan FM, Bickelhaupt DH, Leaf AL (1979) Soil and plant analytical services for tree nurseries. In: Proceedings Northeastern Area Nurseryman's Conference. USDA Forest Service, Northeastern area State and Private Forestry, Broomall, PA: 35-42. https://rngr.net/publications/proceedings/1979/soil-and-plant-analytical-services-for-tree-nurseries/at_download/file
- Sonne E (2006) Greenhouse gas emissions from forestry operations. J Environ Qual 35(4): 1439. <https://dx.doi.org/10.2134/jeq2005.0159>
- South DB (2017) Optimum pH for growing pine seedlings. Tree Planters' Notes 60(2): 49-62. https://rngr.net/publications/tpn/60-2/optimum-ph-for-growing-pine-seedlings/at_download/file
- South DB (2021) Use of boron in conifer nurseries. Reforesta 12:56-97. <https://journal.reforestationchallenges.org/index.php/REFOR/article/download/141/137>

- South DB (2022a) Use of magnesium in bareroot pine nurseries. *Reforesta* 13:7-44. <https://journal.reforestationchallenges.org/index.php/REFOR/article/view/149>
- South DB (2022b) Manganese fertilization in bareroot pine nurseries. *Tree Planters' Notes* 65(2): 60-78.
- South DB, Davey CB (1983) The southern forest nursery soil testing program. R8-TP-4. USDA Forest Service, Southern Region, Atlanta, GA: 140-170. https://rngr.net/publications/1982-southern-nursery-conferences/the-southern-forest-nursery-soil-testing-program/at_download/file
- South DB, Zwolinski JB (1996) Chemicals used in southern forest nurseries. *South J Appl Forest* 20(3): 127-135. <https://doi.org/10.1093/sjaf/20.3.127>
- South DB, Mitchell RJ, Dixon RK, Vedder M (1988) New-ground syndrome: an ectomycorrhizal deficiency in pine nurseries. *South J Appl Forest* 12(4): 234-239. <https://doi.org/10.1093/sjaf/12.4.234>
- South DB, Nadel RL, Enebak SA, Bickerstaff G (2017) Sulfur and lime affect soil pH and nutrients in a sandy *Pinus taeda* nursery. *Reforesta* (4): 12-20. <https://doi.org/10.21750/REFOR.4.02.41>
- South DB, Funk J, Davis CM (2018) Spring fumigation using totally impermeable film may cause ectomycorrhizal deficiencies at sandy loblolly pine nurseries. *Tree Planters' Notes* 61(1): 45-56. https://rngr.net/publications/tpn/61-1/spring-fumigation-using-totally-impermeable-film-may-cause-ectomycorrhizal-deficiencies-at-sandy-loblolly-pine-nurseries/at_download/file
- South DB, Zwolinski JB (1996) Chemicals used in southern forest nurseries. *South J Appl Forest* 20(3): 127-135. <https://doi.org/10.1093/sjaf/20.3.127>
- Starkey T, Enebak S (2012) Foliar nutrient survey of loblolly and longleaf pine seedlings. Research Report 12-02. Auburn University Southern Forest Nursery Management Cooperative, Auburn University, AL: 11 p.
- Steer HB (1915) Some problems of nursery instillation with special reference to white pine in the Northeast. MF thesis, Cornell University, Ithaca. 69 p.
- Steinbeck K (1962) Effects of nutrients on slash pine seedlings grown in different media. Athens, GA: University of Georgia. 67 p. M.S. thesis.
- Steinbeck K (1965) Foliar mineral accumulation by Scotch pine (*Pinus sylvestris* L.) provenances. East Lansing, MI: Michigan State University. 251 p. Ph.D. dissertation. <https://www.proquest.com/docview/302132893?pq-origsite=gscholar&fromopenview=true>
- Steven HM (1928) Nursery investigations. *Forestry* 70: 31-11. <https://www.forestresearch.gov.uk/publications/archive-nursery-investigations/>
- Stoekeler JH (1949) Correction of soil acidity in conifer nurseries. Technical note 319. USDA Forest Service, Lake States Forest Experiment Station, St. Paul, MN: 1 p.
- Stoekeler JH, Jones GW (1957) Forest nursery practice in the Lake States. *Agriculture Handbook* 110. USDA Forest Service, Washington, DC: 124 p. <https://babel.hathitrust.org/cgi/pt?id=uiug.30112019250551&view=1up&seq=1&skin=2021>
- Stone EL (1953) Magnesium deficiency of some northeastern pines. *Soil Sci Soc Am J* 17(3): 297-300. <https://doi.org/10.2136/sssaj1953.03615995001700030029x>
- Stone EL, Hollis CA, Barnard EL (1982) Boron deficiency in a southern pine nursery. *South J Appl Forest* 6 (2): 108-112. <https://doi.org/10.1093/sjaf/6.2.108>
- Su coff EI (1961) Potassium, magnesium, and calcium deficiency symptoms of loblolly and virginia pine seedlings. Station Paper NE-164. USDA Forest Service, Northeastern Forest Experiment Station, Upper Darby, PA: 18 p. https://www.nrs.fs.fed.us/pubs/sp/sp_ne164.pdf
- Su coff EI (1962) Potassium, magnesium, and calcium requirements of Virginia pine. Station Paper NE-169. USDA Forest Service, Northeastern Forest Experiment Station, Upper Darby, PA: 16 p. <https://citeseerx.ist.psu.edu/viewdoc/download?doi=10.1.1.887.3525&rep=rep1&type=pdf>
- Sudworth GB (1900) The forest nursery: collection of tree seeds and propagation of seedlings. Bulletin 29. USDA, Division of Forestry. Washington, DC: 63 p. <https://ir.library.oregonstate.edu/downloads/0z708x74d>
- Sung SS, Black CC, Kormanik TL, Zarnoch SJ, Kormanik PP, Counce PA (1997) Fall nitrogen fertilization and the biology of *Pinus taeda* seedling development. *Can J Forest Res* 27(9): 1406-1412. <https://doi.org/10.1139/x97-112>

- Swan HSD (1970) Relationships between nutrient supply, growth and nutrient concentrations in the foliage of black spruce and jack pine. Woodlands Reports 19. Pulp and paper Research institute of Canada. Pointe Claire. 46 p.
- Switzer GL, Nelson LE (1956) The Effect of fertilization on seedling weight and utilization of N, P, and K by loblolly pine (*Pinus taeda* L.) grown in the nursery. Soil Sci Soc Am J 20(3): 404-408. <https://doi.org/10.2136/sssaj1956.03615995002000030029x>
- Tanaka H, Yatazawa M, Iyer JG (1967) Supply of trace elements in nursery soils of Wisconsin. Soil Sci Plant Nutr 13(1): 31-35. <https://doi.org/10.1080/00380768.1967.10431970>
- Tillotson CR (1917) Nursery practice on the national forests. Bulletin 479. USDA. Washington, DC: 86 p. <https://babel.hathitrust.org/cgi/pt?id=uiug.30112019242087&view=1up&seq=1&skin=2021>
- Timmer VR, Armstrong G (1987) Diagnosing nutritional status of containerized tree seedlings: comparative plant analyses1. Soil Sci Soc Am J 51(4): 1082-1086. <https://doi.org/10.2136/sssaj1987.03615995005100040048x>
- Timmer VR, Parton WJ (1984) Optimum nutrient levels in a container growing medium determined by a saturated aqueous extract. Commun Soil Sci Plant 15(6): 607-618. <https://doi.org/10.1080/00103628409367502>
- Toca A, Oliet JA, Villar-Salvador P, Maroto J, Jacobs DF (2018) Species ecology determines the role of nitrogen nutrition in the frost tolerance of pine seedlings. Tree physiol 38(1): 96-108. <https://doi.org/10.1093/treephys/tpx165>
- Talkner U, Riek W, Dammann I, Kohler M, Göttlein A, Mellert KH, Meiwes KJ (2019) Nutritional status of major forest tree species in Germany. In: Status and Dynamics of Forests in Germany. Ecological Studies 237: 261-293. https://link.springer.com/chapter/10.1007/978-3-030-15734-0_9
- Toumey JW (1916) Seeding and planting: a manual for the guidance of forestry students, foresters, nurserymen, forest owners, and farmers. John Wiley & Sons, NY. <https://digital.library.cornell.edu/catalog/chla2731710>
- Truman RA (1972) The visual diagnosis of mineral deficiencies in *Pinus radiata*, *P. elliotii* and *P. taeda* seedlings. For. Comm. N.S.W. Tech. Pap. No. 19. 18 p. https://www.dpi.nsw.gov.au/_data/assets/pdf_file/0011/389909/The-Detection-of-Mineral-Deficiencies-in-Pinus-Elliottii,-Pinus-Radiata-and-Pinus-Taeda-by-Visual-Means.pdf
- van den Burg J (1990) Foliar analysis for determination of tree nutrient status: a compilation of literature data: 2: Literature 1985-1989. "De Dorschkamp" Institute for Forestry and Urban Ecology, Wageningen. <https://library.wur.nl/WebQuery/wurpubs/527874>
- van den Driessche R (1963) Nursery experiments with Douglas fir. The Commonwealth Forestry Review 42(3): 242-254. <https://www.jstor.org/stable/42603050>
- van den Driessche R (1984) Soil fertility in forest nurseries. In: Duryea ML, Landis TD (eds) Forest Nursery Manual. MartinusNijhoff/Junk Publishers, The Hague, The Netherlands: 63-74. <https://rngr.net/publications/nursery-manuals/fnm/Chapter%207/>
- van den Driessche R (1991) Effects of nutrients on stock performance in the forest. In: van den Driessche (ed) Mineral Nutrition of Conifer Seedlings, CRC Press, Boca Raton: 229-260.
- VandeHey JMV (2007) Production of conifer bareroot seedlings using controlled release fertilizer. Native Plants Journal 8(3): 288-293. <http://npj.uwpress.org/content/8/3/288.short>
- van Lear DH, Smith WH (1972) Relationships between macro-and micronutrient nutrition of slash pine on three coastal plain soils. Plant Soil 36(1-3): 331-347. <https://doi.org/10.1007/BF01373488>
- Vettorazzo SC; Couto HTZ (1997) Padronização de amostragem para diagnose nutricional e variações nas concentrações de nutrientes nas acículas de *Pinus taeda* L. I. Macronutrientes. Revista Brasileira de Ciência do Solo 21: 51-58. <https://pascal-francis.inist.fr/vibad/index.php?action=getRecordDetail&idt=2715028>
- Villarrubia JM (1980) Effect of nitrogen rate and source on growth and performance of *Liquidambar styraciflua* (sweetgum) and *Fraxinus pennsylvanica* (green ash) in a Virginia nursery. PhD thesis, North Carolina State University, Raleigh. 91 p. <https://catalog.lib.ncsu.edu/catalog/NCSU567792>
- Voigt GK (1955) The effect of applied fungicides, herbicides, and insecticides on the content of nutrient elements in tissue of coniferous seedlings. Soil Sci Soc Am J 19(2): 237-239. <https://doi.org/10.2136/sssaj1955.03615995001900020033x>

- Voigt GK, Stoeckeler JH, Wilde SA (1958) Response of coniferous seedlings to soil applications of calcium and magnesium fertilizers. *Soil Sci Soc Am J* 22(4): 343-345. <https://access-onlinelibrary-wiley-com.spot.lib.auburn.edu/doi/abs/10.2136/sssaj1958.03615995002200040022x>
- Wahlenberg WG (1930) Experiments in use of fertilizers in growing forest planting material at Savenac Nursery. Circular 125. USDA, Washington, DC: 38 p. <https://openlibrary.org/books/OL19658308M>
- Wakeley PC (1935) Artificial reforestation in the southern pine region. Technical Bulletin 492. USDA, Washington, DC: 114 p. <https://ageconsearch.umn.edu/record/164666/files/tb492.pdf>
- Walker RF, McLaughlin SB (1997) Effects of acidic precipitation and ectomycorrhizal inoculation on growth, mineral nutrition, and xylem water potential of juvenile loblolly pine and white oak. *J Sustain Forest* 5(3-4): 27-49. https://doi.org/10.1300/J091v05n03_03
- Wall MM (1994) Influence of fertilization on nutrient status and size of bare-root *Pinus taeda* L. seedlings. MS thesis, Texas A&M University, College Station. 98 p. <https://hdl.handle.net/1969.1/ETD-TAMU-1994-THESIS-W1875>
- Wallander H, Wickman T, Jacks G (1997) Apatite as a P source in mycorrhizal and non-mycorrhizal *Pinus sylvestris* seedlings. *Plant and Soil* 196(1): 123-131. <https://doi.org/10.1023/A:1004230525164>
- Weetman GF, Algar D (1974) Jack pine nitrogen fertilization and nutrition studies: three year results. *Can J Forest Res* 4(3): 381-398. <https://doi.org/10.1139/x74-056>
- Wells CG, Crutchfield DM, Berenyl NM, Davey CB (1973) Soil and foliar guidelines for phosphorus fertilization of loblolly pine. USDA Forest Service, Southeastern Forest Experiment Station, Asheville, NC: 15 p. https://www.srs.fs.usda.gov/pubs/rp/rp_se110.pdf
- White EH, Comerford NB, Bickelhaupt DH (1980) Interpretation of nursery soil and seedling analysis to benefit nursery soil management. In: Proceedings of the North American Forest Tree Nursery Soils Workshop. Syracuse, NY: 269-287. <https://babel.hathitrust.org/cgi/pt?id=pst.000007190872&view=1up&seq=9&skin=2021>
- White PJ (2015) Calcium. In Barker AV, Pilbeam DJ (eds), Handbook of plant nutrition. CRC press. 165-198. <https://www.researchgate.net/file/PostFileLoader.html?id=5145f99ce24a46c827000013&assetKey=AS:271834098929664@1441821709168>
- Wilde SA (1938) Soil fertility standards for growing northern conifers in forest nurseries. *J Agric Res* 57: 945-952. <https://naldc.nal.usda.gov/download/IND43969114/PDF>
- Wilde SA (1940) What the Department has learned in nursery soil management. *Wisconsin Conservation Bulletin* 5(10): 3-6. <https://babel.hathitrust.org/cgi/pt?id=uc1.b3065799&view=1up&seq=777&skin=2021&q1=wilde>
- Wilde SA (1942) Forest soils origin, properties, relation to vegetation, and silvicultural management. Kramer Business Service, Madison. <https://babel.hathitrust.org/cgi/pt?id=mdp.39015004577030&view=1up&seq=2&skin=2021>
- Wilde SA (1946) Forests soils and forests growth. *Chronica Botanica Company*, Waltham. <https://reader.library.cornell.edu/docviewer/digital?id=chla5566828>
- Wilde SA (1958) Forest soils, their properties and relation to silviculture. Ronald Press, NY. <https://doi.org/10.2134/agronj1959.00021962005100100030x>
- Wilde SA, Kopitke JC (1940) Base exchange properties of nursery soils and the application of potash fertilizers. *J Forest* 38(4): 330-332. <https://doi.org/10.1093/jof/38.4.330>
- Will GM (1961) Magnesium deficiency in pine seedlings growing in pumice soil nurseries. *New Zealand Journal of Agricultural Research* 4(1-2): 151-160. <https://www.tandfonline.com/doi/pdf/10.1080/00288233.1961.10419929>
- Will GM (1985) Nutrient deficiencies and fertilizer use in New Zealand exotic forests. *NZ For Res Inst Bull* 97: 53 p. <https://scion.contentdm.oclc.org/digital/api/collection/p20044coll6/id/264/download>
- Will GM, Knight PJ (1968) Pumice soils as a medium for tree growth: pot trial evaluation of nutrient supply. *New Zealand Journal of Forestry* 13(1): 50-65. http://nzjf.org.nz/free_issues/NZJF13_1_1968/C3FD6A3B-07E4-4B10-81C0-C8214E135DD2.pdf

- Woessner RA, Davey CB, Crabtree BE, Gregory JD (1975) Nutrient content of the aboveground tissue of 12-week-old loblolly pine intraprovenance and interprovenance crosses. *Canadian Journal of Forest Research* 5(4): 592-598. <https://doi.org/10.1139/x75-086>
- Wood T, Bormann FH (1977) Short-term effects of a simulated acid rain upon the growth and nutrient relations of *Pinus strobus*, L. *Water, Air, and Soil Pollution* 7(4): 479-488. <https://doi.org/10.1007/BF00285546>
- Woodwell GM (1958) Factors controlling growth of pond pine seedlings in organic soils of the Carolinas. *Ecol Monogr* 28(3): 220-236. <https://doi.org/10.2307/1942241>
- Woollons RC, Crane WJB, Snowdon P (1995) Responses to nitrogen, phosphorus and sulphur applications to a *Pinus radiata* stand in the Tumut region, New South Wales. *Australian Forestry* 58(3): 135-141. <https://doi.org/10.1080/00049158.1995.10674656>
- Youngberg CT (1958) The uptake of nutrients by western conifers in forest nurseries. *J Forest* 56(5): 337-340. <https://doi.org/10.1093/jof/56.5.337>
- Youngberg CT (1984) Soil tissue analysis: tools for maintaining soil fertility. In: Duryea ML, Landis TD (eds.) *Forest nursery manual*. MartinusNijhoff/Junk Publishers. The Hague, Netherlands: 75-80. https://link.springer.com/content/pdf/10.1007%2F978-94-009-6110-4_7.pdf
- Zhang J, George E (2010) Effect of the ectomycorrhizal fungus *Paxillus involutus* on growth and cation (potassium, calcium, and magnesium) nutrition of *Pinus sylvestris* L. in semi-hydroponic culture. *J Plant Nutr* 33(5): 736-751. <https://doi.org/10.1080/01904160903575956>
- Zöttl HW (1973) Diagnosis of nutritional disturbances in forest stands. Presented to. *FAO/IUFRO International symposium on forest fertilization, Paris, 75-95.*